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(54) Title: CARBOHYDRATE CROSSLINKED GLYCOPROTEIN CRYSTALS			
(57) Abstract			
<p>The present invention relates to the field of carbohydrate crosslinked glycoprotein crystals. Advantageously, such crosslinked glycoprotein crystals display stability to harsh environmental conditions, while maintaining the structural and functional integrity of the glycoprotein backbone. According to one embodiment, this invention relates to methods for concentrating proteins that have been modified by carbohydrates and for releasing their activity at controlled rates. This invention also provides methods for producing carbohydrate crosslinked glycoprotein crystals and methods for using them in pharmaceutical formulations, vaccines, immunotherapeutics, personal care compositions, including cosmetics, veterinary pharmaceutical compositions and vaccines, foods, feeds, diagnostics, cleaning agents, including detergents and decontamination formulations. The physical and chemical characteristics of carbohydrate crosslinked glycoprotein crystals render them particularly useful as sorbents for separations, such as chiral chromatography, or affinity chromatography – which are based on specific interactions between the active binding site of the glycoprotein component of the crystals and the substance or molecule of interest. Such characteristics also render carbohydrate crosslinked glycoprotein crystals useful as catalytic and binding components for the production of biosensing devices.</p>			

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CARBOHYDRATE CROSSLINKED GLYCOPROTEIN CRYSTALSTECHNICAL FIELD OF THE INVENTION

The present invention relates to the field of carbohydrate crosslinked glycoprotein crystals.

5 Advantageously, such crosslinked glycoprotein crystals display stability to harsh environmental conditions, while maintaining the structural and functional integrity of the glycoprotein backbone. According to one embodiment, this invention relates to methods for

10 concentrating proteins that have been modified by carbohydrates and for releasing their activity at controlled rates. This invention also provides methods for producing carbohydrate crosslinked glycoprotein crystals and methods for using them in pharmaceutical formulations, vaccines, immunotherapeutics, personal care compositions, including cosmetics, veterinary pharmaceutical compositions and vaccines, foods, feeds, diagnostics, cleaning agents, including detergents and decontamination formulations. The physical and

15 20 chemical characteristics of carbohydrate crosslinked glycoprotein crystals render them particularly useful as sorbents for separations, such as chiral chromatography, or affinity chromatography -- which are based on specific interactions between the active

- 2 -

binding site of the glycoprotein component of the crystals and the substance or molecule of interest. Such characteristics also render carbohydrate crosslinked glycoprotein crystals useful as catalytic 5 and binding components for the production of biosensing devices.

BACKGROUND OF THE INVENTION

Many proteins associated with the external surfaces of cell membranes or actively secreted from 10 cells are commonly modified by the addition of one or more carbohydrate units to the side chains of particular amino acids [R.D. Marshall, Ann. Rev. Biochem., 41, pp. 673-702 (1972)]. Such proteins, known as glycoproteins, display the properties of 15 proteins in general, as well as properties typical of the attached carbohydrate. The carbohydrate monomers typically attached to glycoproteins include galactose, mannose, glucose, N-acetylglucosamine, N-acetylgalactosamine, fucose, xylose, sialic acid and 20 others. The carbohydrate units are usually attached through the hydroxyl groups of serine and threonine side chains, or the amide nitrogen atom of asparagine side chains. The carbohydrate side chains are arranged in a variety of chain lengths and branching patterns 25 [P.V. Wagh and O.P. Bahl, Crit. Rev. Biochem., 10, pp. 307-77 (1981)].

Glycoproteins exhibit a range of protein functions, including catalysis of chemical transformations, proteolysis of proteins, binding of 30 ligands and transport of ligands to and across membranes. Additionally, glycoproteins frequently perform functions associated with cellular

- 3 -

communication, including protein-protein recognition, protein-carbohydrate recognition, protein-DNA recognition, pathogen recognition by antibodies, antigen presentation by CD4 and CD8 membrane glycoproteins, and targeting of proteins to specific locations.

Glucose oxidase exemplifies an enzyme glycoprotein that catalyzes the oxidation of  $\beta$ -D-glucose to D-glucono-1,5-lactone. The reaction consumes one mole of oxygen and produces one mole of hydrogen peroxide per mole of glucose. The active glycoprotein glucose oxidase, or  $\beta$ -D-glucose:oxygen 1-oxidoreductase [enzyme commission number 1.1.3.4] forms a dimer with a molecular weight of 150-180 kDa. Each monomer consists of 583 amino acids residues, one co-factor molecule of flavin adenine dinucleotide (FAD) and has a carbohydrate content of approximately 16% by weight. The three dimensional structure of the deglycosylated protein has been determined by X-ray crystallography [H.J. Hecht, H.M. Kalisz, J. Hendle, R.D. Schmid and D. Schomburg, J. Mol. Biol., 229, pp.153-72 (1993)]. Due to the importance of the quantitative determination of glucose in medicine and industry, glucose oxidase is considered a prime candidate for the development of biosensors [H.J. Hecht, D. Schomburg, H.M. Kalisz and R.D. Schmid, Biosensors and Bioelectronics, 8, pp. 197-203 (1993)]. Glucose oxidase may be advantageously used in the food, drug and cosmetics industries, because of its ability to interconvert oxygen and hydrogen peroxide. Commercially available glucose oxidase is usually isolated from *Aspergillus niger*.

- 4 -

The stereoselectivity and specific activity of enzymatic glycoproteins may be exploited for use in industrial syntheses. For example, glutaraldehyde crosslinked crystals of Lipase from *Candida rugosa* may 5 be used to synthesize optically pure compounds [J.J. Lalonde, C. Govardhan, N. Khalaf, A.G. Martinez, K. Visuri and A.L. Margolin, J. Am. Chem. Soc., 117, (26) pp. 6845-52, (1995)].

In infectious diseases, glycoproteins are 10 involved in the initiation and maintenance of infection, as well as host humoral and cellular immune responses against infection. The surface proteins of many viruses are glycoproteins. Examples of such glycoproteins include, for example, gp120 and gp41 of 15 the Human Immunodeficiency Virus (HIV), which causes AIDS [H. Geyer, C. Holschbach, G. Hunsmann and J. Schneider, J. Biol. Chem., 263 (24), pp. 11760-67, (1988)] and the hemagglutinin (HA) and neuraminidase (NA) of Influenza Virus, which causes Flu.

Viral receptors, the cellular proteins 20 recognized by invading viruses, are found on the surface of cells and therefore are frequently either glycoproteins or carbohydrates. The CD4 molecule, which is recognized by HIV-1 gp120, is a glycoprotein. 25 Similarly, Influenza virus hemagglutinin binds to terminal N-acetylneurameric acid residues of sialoglycoproteins and enters the cell through receptor mediated endocytosis [J. White, M. Kielian and A. Helenius, Quart. Rev. of Biophys., 16, pp. 151-95, 30 (1983)].

In spite of the tremendous medical, chemical, pharmaceutical and industrial potential of glycoproteins, their development has, in many

- 5 -

instances, lagged far behind that of unglycosylated proteins.

As compared with unglycosylated proteins, the frequent association of glycoproteins with biological membranes and other membrane proteins, render glycoproteins significantly more difficult to purify and utilize for medical and industrial processes. The use of glycoproteins faces additional barriers because relatively little is known about their three-dimensional structure and the requirements for stabilization when faced with harsh environments. However, due to the specialized functions of glycoproteins, many benefits can be realized by overcoming the barriers to widespread large scale use of glycoproteins in industrial, chemical and medical applications.

One unique approach to overcoming barriers to the widespread use of proteins generally is crosslinked enzyme crystal ("CLEC™") technology [N.L. St. Clair and M.A. Navia, J. Am. Chem. Soc., 114, pp. 4314-16 (1992)]. Crosslinked enzyme crystals retain their activity in environments that are normally incompatible with enzyme function. Such environments include prolonged exposure to proteases and other protein digestion agents, high temperature or extreme pH and organic solvents. In such environments, crosslinked enzyme crystals remain insoluble and stable.

One physical result of "CLEC™" technology is that the surface exposed amino acid side chains of the protein, in the crystal lattice, are covalently modified with the crosslinking agents. This modification stabilizes the crystal lattice, at the same time altering elements of the surface structure to gain the benefits of stabilization. For most

- 6 -

applications involving protein crystals, any potential limitation resulting from minor surface modifications is overcome by the gains in stability achieved in the crystals. However, applications involving vaccines,  
5 and immunotherapeutics using glycoproteins, often require that the surface exposed protein structures, known as epitopes, precisely maintain the original structure. This may require stabilization without chemical modification of the amino acid side chains or  
10 perhaps with minor levels of chemical modification of the amino acid side chains.

DISCLOSURE OF THE INVENTION

The present invention specifically addresses the problems of stabilization of glycoproteins via  
15 crystallization and crosslinking for their use in industrial, chemical, and medical applications. Thus, the present invention relates to methods for crystallization of glycoproteins and for their subsequent stabilization through carbohydrate-to-  
20 carbohydrate crosslinking within the crystal lattice to produce carbohydrate crosslinked glycoprotein crystals.

Crosslinked glycoprotein crystals according to this invention may be produced by crosslinking glycoprotein crystals through one or more carbohydrate  
25 moieties on the glycoprotein or through both one or more carbohydrate moieties on the glycoprotein and one or more of the amino acid side chain functional groups in the glycoprotein.

The carbohydrate crosslinked glycoprotein  
30 crystals of this invention are useful in vaccines, pharmaceuticals and immunotherapeutic formulations for delivery of active, non-denatured, glycoproteins. They

- 7 -

are also useful in biosensing devices and in methods, devices and systems for separating molecules of interest from mixtures or crude preparations thereof.

The present invention also includes methods  
5 for crystallizing glycoproteins in the absence of the potentially denaturing effects of chemical deglycosylation.

#### DETAILED DESCRIPTION OF THE INVENTION

In order that the invention herein described  
10 may be more fully understood, the following detailed description is set forth. In the description, the following terms or phrases are employed:

Aqueous-organic solvent mixture -- a mixture comprising n% organic solvent, where n is between 1 and  
15 99 and m% aqueous, where m is 100-n.

Catalytically effective amount -- an amount of carbohydrate crosslinked glycoprotein crystals of this invention which is effective to treat, protect, repair or detoxify the area to which they are applied  
20 over some period of time.

Change in chemical composition -- any change in the chemical components of the environment surrounding carbohydrate crosslinked glycoprotein crystals that affects the environment or the  
25 crosslinker, including addition of chemical reagents, solvent composition changes, chemical changes induced by application of energy in the form of light, microwave, or radiation to the environment, chemical events and parameters that affect the crosslinker and combinations thereof.

Change in shear force acting upon the crystals -- any change in factors of the environment

- 8 -

surrounding carbohydrate crosslinked glycoprotein crystals under conditions of use, such as, changes in mechanical pressure, both positive and negative, revolution stirring, centrifugation, tumbling,

5 mechanical agitation and filtration pumping.

Controlled dissolution -- dissolution of carbohydrate crosslinked glycoprotein crystals or release of the protein constituent of said crystals that is (1) triggered by a change in the environment 10 surrounding said crystals, said change being selected from the group consisting of change in temperature, change in pH, change in chemical composition, change from concentrate to dilute form, change in shear force acting upon the crystals and combinations thereof and 15 (2) controlled by a factor selected from the group consisting of the degree of crosslinking of said carbohydrate crosslinked glycoprotein crystals, the length of time of exposure of glycoprotein crystals to the crosslinking agent, the rate of addition of 20 crosslinking agent to said glycoprotein crystals, the nature of the crosslinker, the chain length of the crosslinker, the surface area of said carbohydrate crosslinked glycoprotein crystals, the size of said carbohydrate crosslinked glycoprotein crystals, the 25 shape of said carbohydrate crosslinked glycoprotein crystals and combinations thereof.

Formulations for decontamination --

formulations selected from the group consisting of: formulations for decontamination of chemical wastes, 30 herbicides, insecticides, pesticides, environmental hazards.

Glycoprotein -- a protein or peptide covalently linked to carbohydrate. The carbohydrate may be monomeric or composed of oligosaccharides.

- 9 -

Glycoprotein activity -- an activity selected from the group consisting of binding, catalysis, signalling, transport, or other activities which induce a functional response within the environment in which 5 the glycoprotein is used, such as induction of immune response and immunogenicity, or combinations thereof.

Glycoprotein activity release rate -- the quantity of glycoprotein dissolved per unit time.

Insoluble and stable form of a glycoprotein  
10 -- a form of a glycoprotein which is insoluble in aqueous solvents, organic solvents or aqueous-organic solvent mixtures and which displays greater stability than the soluble form of the glycoprotein. According to an alternate embodiment of this invention, the  
15 phrase "insoluble and stable form of a glycoprotein" may be a protein which is insoluble in dry formulations but soluble in wet formulations. In any embodiment, carbohydrate crosslinked glycoprotein crystals may be active in insoluble form. And in one embodiment,  
20 carbohydrate crosslinked glycoprotein crystals according to this invention may be active in insoluble form, then dissolve or are removed or digested once their function is complete.

Immunotherapeutic -- a protein or  
25 glycoprotein derived from a tumor cell with a protein activity of inducing protective immunity to said tumor. A protein or glycoprotein cytokine which stimulates the immune system to reduce or eliminate said tumor.

Organic solvents -- any solvent of non-aqueous origin.

Pharmaceutically effective amount -- an amount of carbohydrate crosslinked glycoprotein crystals which is effective to treat a condition in an

- 10 -

individual to whom they are administered over some period of time.

Prophylactically effective amount -- an amount of carbohydrate crosslinked glycoprotein 5 crystals which is effective to prevent a condition in an individual to whom they are administered over some period of time.

Protein -- any peptide having a tertiary structure or any protein.

10           Separation -- Separation of a substance from a mixture of two or more different substances or two or more forms of the same substance. According to another embodiment of this invention, "separation" is defined as purification of a substance from a crude form 15 thereof. Separation may be carried out by any means including, for example, chromatography, membrane separation, filtration and electrophoresis.

20           Therapeutic glycoprotein -- A glycoprotein which is administered to a patient in a conventional pharmaceutical formulation and manner. Therapeutic glycoproteins include, for example, hormones, enzymes, antibodies, viral receptors, T-cell receptors, chemokines, chemokine receptors, MHC molecules, tumor antigens, mucins, inhibitors, growth factors, trophic 25 factors, cytokines, lymphokines, toxoids, nerve growth hormones, blood clotting factors, adhesion molecules, multidrug resistance proteins, adenylyl cyclases and bone morphogenic proteins.

30           Vaccine antigen -- a protein or glycoprotein derived from an infectious agent such as a virus, parasite, or tumor antigen. The protein activity of such vaccine antigens is to induce protective immunity against the infectious agent.

- 11 -

As a result of their crystalline nature, the carbohydrate crosslinked glycoprotein crystals of this invention achieve uniformity across the entire crosslinked crystal volume. This uniformity is  
5 maintained by the intermolecular contacts and chemical crosslinks between the carbohydrates attached to adjacent protein molecules constituting the crystal lattice, even when the crystals are exposed to buffers, organic or mixed aqueous-organic solvents and  
10 adjuvants. In such media, the glycoprotein molecules maintain a uniform distance from each other, forming well-defined stable pores within the carbohydrate crosslinked glycoprotein crystals that facilitate access of substrate or ligand to the glycoproteins, as  
15 well as removal of product.

The methods of this invention achieve stabilization of the crystal lattice by either exclusive crosslinking of attached carbohydrates of glycoproteins or a combination of carbohydrate and  
20 amino acid side chain crosslinking. In such carbohydrate crosslinked glycoprotein crystals, the lattice interactions, when fixed by chemical crosslinks, are particularly important in providing stability and preventing denaturation, especially in  
25 storage, under conditions including harsh environments created by components of compositions in which the crystals are used. The uniformity across crystal volume and enhanced stability of the constituent glycoproteins in carbohydrate crosslinked glycoprotein  
30 crystals creates novel opportunities for the use of glycoprotein vaccines, biosensors, and catalysis in harsh conditions, such as elevated temperature, and aqueous, organic or near-anhydrous solvents, as well as mixtures of these. Glycoprotein crystals may also be

- 12 -

crosslinked in such a way that they dissolve or release their protein activity upon exposure to a trigger in their environment encountered under conditions of use. Thus, they may be substantially insoluble and stable in  
5 a composition under storage conditions and substantially soluble and active under conditions of use of said composition.

The methods of this invention advantageously accomplish the crystallization of glycoproteins, under  
10 large scale conditions, without the need for cumbersome and potentially denaturing effects of chemical deglycosylation. As a result, the carbohydrate moieties attached to amino acid side chains available for chemical crosslinking are maintained, while  
15 preserving those amino acid side chains not attached to carbohydrate in their unmodified form.

For those carbohydrate crosslinked glycoprotein crystals according to this invention which are enzymes, the entire crystal consists of active  
20 enzyme (and not inactive carrier). The specific activity per mg of immobilized protein product in carbohydrate crosslinked glycoprotein crystals is typically at least about 2 times higher than in conventionally immobilized proteins or catalysts,  
25 ranging from about 2 to 100 times higher. Such high glycoprotein densities are particularly useful in biosensor, analytical and other applications requiring large amounts of protein in small volumes.

Carbohydrate crosslinked glycoprotein  
30 crystals according to this invention offer several advantages over conventional protein/glycoprotein immobilization methods. For example, the crosslinked crystal matrix provides its own support. Expensive carrier beads, glasses, gels, or films are not required

- 13 -

in order to tie down the enzyme catalyst, as they are in presently available immobilization methods. As a result, the concentration of glycoprotein is close to the theoretical packing limit that can be achieved for 5 molecules of a given size, greatly exceeding densities achievable even in concentrated solutions.

In addition to their activity, carbohydrate crosslinked glycoprotein crystals according to this invention are particularly stable and insoluble under 10 storage conditions, including the attendant storage temperature, storage pH, storage time, storage concentrate form, storage involving little or no shear force acting upon the crystals, or combinations thereof.

15 In addition to their stability under storage conditions, carbohydrate crosslinked glycoprotein crystals are particularly stable, at least about 2 times as stable, to thermal denaturation, digestion with proteases, for example pronase, and in mixed 20 water:organic solvent mixtures. Specifically, the carbohydrate crosslinked glycoprotein crystals of this invention are between about 2 and 100 times as stable to thermal denaturation, between about 2 and 10,000 times more resistant to pronase digestion, and between 25 about 2 and 1,000 times more stable to inactivation in 50% ethanol, than the soluble form of the enzyme.

The rate of dissolution of carbohydrate crosslinked glycoprotein crystals can be controlled by manipulating the conditions and extent of crosslinking. 30 Controlled dissolution carbohydrate crosslinked glycoprotein crystals are slowly soluble and active under conditions of use, including conditions involving change in temperature, change in pH, change in chemical composition, change from concentrate to dilute form,

- 14 -

change in shear force acting upon the crystals and combinations thereof. However such carbohydrate crosslinked glycoprotein crystals are insoluble and stable under storage conditions. Such properties make  
5 the carbohydrate crosslinked glycoprotein crystals of this invention particularly useful for delivery of pharmaceuticals, therapeutic glycoproteins, personal care agents or compositions, including cosmetics, vaccines, veterinary compositions, foods, feeds,  
10 diagnostics, cleaning agents, including detergents and formulations for decontamination.

According to one embodiment, the carbohydrate crosslinked glycoprotein crystals of this invention are characterized by a half-life of activity under storage  
15 conditions which is greater than at least about 2 times that of the soluble form of the glycoprotein that is crystallized to form the crystals that are crosslinked, as well as activity similar to that of the soluble form of the glycoprotein under conditions of use.

20 Advantageously however, the carbohydrate crosslinked glycoprotein crystals of this invention display improved stability under storage conditions, as compared to their soluble or uncrosslinked crystallized counterpart glycoproteins.

25 Thus, carbohydrate crosslinked glycoprotein crystals according to this invention may be advantageously used instead of conventional soluble or immobilized proteins in pharmaceuticals, veterinary compounds, personal care compositions, including  
30 cosmetics, foods, feeds, vaccines, pulp, paper and textile processing, diagnostics, cleaning agents, including detergents and formulations for decontamination.

- 15 -

The carbohydrate crosslinked glycoprotein crystals of this invention are particularly advantageous because they are stable in harsh environments imposed by the formulations or 5 compositions in which they are employed or conditions of their storage. At the same time, these carbohydrate crosslinked glycoprotein crystals are capable of (1) change to soluble and active form (an active form including, in one embodiment of this invention, a form 10 which is active against macromolecular substrates) or (2) controlled dissolution or release of their activity when exposed to one or more triggers in their environment. Such triggers may be selected from the group consisting of change in temperature, change in 15 pH, change in chemical composition, change from concentrate to dilute form, change in shear force acting upon the crystals and combinations thereof. Controlled dissolution or release of activity of carbohydrate crosslinked glycoprotein crystals 20 according to this invention may also be triggered over a change in time.

Specific examples of such triggers include an increase or decrease in temperature, for example, an increase in temperature from a low temperature between 25 about 0°C and about 20°C to a high temperature between about 25°C and about 70°C. Other triggers include a change from acidic pH to basic pH and a change from basic pH to acidic pH. Examples of triggers of change from concentrate to dilute form include, for example, a 30 change in solute concentration, a change in concentration of all solutes from about 2-fold to about 10,000-fold, a change in concentration of all solutes from about 2-fold to about 700-fold, an increase or decrease in salt concentration, an increase or decrease

- 16 -

in water concentration, an increase or decrease in organic solvent concentration, a decrease in protein concentration and a decrease in detergent concentration.

5           Additional triggers involve changes in chemical composition of the environment surrounding the carbohydrate crosslinked glycoprotein crystals that affect the environment or the crosslinker itself. Such changes include, for example, addition of chemical  
10 reagents, increase or decrease in organic solvent concentration, chemical events that affect the crosslinker, chemical changes induced by application of energy, including light, microwave or radiation. As explained above, any of these triggers may act in  
15 combination or in sequence with one or more of the other triggers.

The glycoprotein constituent of the carbohydrate crosslinked glycoprotein crystals of this invention may be any glycoprotein, including for  
20 example, hormones, enzymes, antibodies, viral receptors, viral surface glycoproteins, parasite glycoproteins, parasite receptors, T-cell receptors, MHC molecules, immune modifiers, tumor antigens, mucins, inhibitors, growth factors, trophic factors,  
25 cytokines, lymphokines, cytokines, toxoids, nerve growth hormones, blood clotting factors, adhesion molecules, multidrug resistance proteins, adenylate cyclases, bone morphogenic proteins and lectins.

Also included among the glycoproteins are the  
30 hormones and cytokines. Examples of hormones include follicle stimulating hormone, human chorionic gonadotropin, luteinizing hormone, thyrotrophin and ovine, bovine, porcine, murine and rat alleles of these hormones. Examples of cytokine glycoproteins include

- 17 -

$\alpha$ -interferon, lymphotoxin, and interleukin-2. Also included are glycoprotein tumor-associated antigens, for example, carcinoembryonic antigen (CEA), human mucins, her-2/neu, and prostate-specific antigen (PSA)

5 [R.A. Henderson and O.J. Finn, Advances in Immunology, 62, pp. 217-56 (1996)].

Alternatively, the glycoprotein constituent may be selected from personal care glycoproteins, including cosmetic glycoproteins, veterinary

10 glycoproteins, food glycoproteins, feed glycoproteins, diagnostic glycoproteins, glycoproteins used in chemical reactions, glycoproteins used in industrial methods, cleaning agent glycoproteins, including detergent glycoproteins, and decontamination

15 glycoproteins. Included among such glycoproteins are enzymes, such as, for example, hydrolases, transferases, isomerases, lyases, ligases, transferases and oxidoreductases. Examples of hydrolases include lipase, cholinesterase, alkaline phosphatase,  $\beta$ -amylase

20 deoxyribonuclease, glucoamylase A and B,  $\alpha$ -galactosidase I and II,  $\beta$ -fructofuranosidase,  $\beta$ -glucuronidase, N-acetyl- $\beta$ -glucosaminidase, hyaluronidase, oxytocinase, kallikrein, bromelain, enterokinase, proteinase a, b, and c, pepsinogen and

25 pepsin. Examples of oxidoreductases include glucose oxidase, peroxidase and chloroperoxidase. Examples of transferases include  $\gamma$ -glutamyltranspeptidase and ribonuclease.

In one embodiment of this invention,

30 carbohydrate crosslinked glycoprotein crystals are produced by a method comprising at least one crosslinking reaction in which at least one carbohydrate moiety attached to the glycoprotein acts

- 18 -

as or functions as the substrate for the crosslinking reaction. Multiple carbohydrate crosslinking reactions can be performed to modify further the characteristics of the carbohydrate crosslinked glycoprotein crystals.

5       The carbohydrate crosslinked glycoprotein crystals of this invention may be prepared using crosslinking reagents which are multifunctional or bifunctional agents. Such agents include the diamine group of compounds, such as, for example,

10 hexamethylenediamine, diaminooctane, ethylenediamine, 4-(4-N-Maleimidophenyl)butyric acid hydrazide·HCl (MPBH), 4-(N-Maleimidomethyl)cyclohexane-1-carboxyhydrazide·HCl ( $M_2C_2H$ ), and 3-(2-Pyridyldithio)propionyl hydrazide (PDPH) and other amine alkenes.

15       In one embodiment of this invention, carbohydrate crosslinked glycoprotein crystals are produced by a method including at least one crosslinking reaction in which at least one carbohydrate moiety attached to the glycoprotein acts

20 or functions as the substrate for the crosslinking reaction. One method of producing carbohydrate crosslinked glycoprotein crystals comprises an initial oxidation of the carbohydrate moieties, followed by crosslinking with at least a multifunctional reagent

25 such as a diamine, followed by a reduction reaction using for example,  $NaBH_4$ . Alternatively, additional carbohydrate crosslinking reactions can be carried out as described but using different crosslinking reagents.

30       In another embodiment of this invention, carbohydrate crosslinked glycoprotein crystals are produced by a method including an initial crosslinking reaction in which one or more amino acid side chain functional groups serve as substrate for a

- 19 -

multifunctional crosslinking reagent, such as glutaraldehyde. The first crosslinking reaction is followed by additional crosslinking reactions in which at least one involves crosslinking through one or more 5 carbohydrate moieties and using, for example, a diamine crosslinking reagent.

In another embodiment of this invention, at least one crosslinking reaction is performed, in addition to the crosslinking reaction in which at least 10 one oxidized carbohydrate moiety attached to the glycoprotein acts as or functions as the substrate for the crosslinking reaction, in which the amino acid side chain functional groups act as or function as a substrate for the crosslinking reaction. This 15 additional linking can be achieved using one or a combination of a wide variety of multifunctional crosslinking reagents, at the same time (in parallel) or in sequence. Such multifunctional reagents include bifunctional reagents. Examples of such crosslinking 20 agents are glutaraldehyde, succinaldehyde, octanodialdehyde and glyoxal. Additional multifunctional crosslinking agents include halo-triazines, e.g., cyanuric chloride; halo-pyrimidines, e.g., 2,4,6-trichloro/bromo-pyrimidine; anhydrides or 25 halides of aliphatic or aromatic mono- or di-carboxylic acids, e.g., maleic anhydride, (meth)acryloyl chloride, chloroacetyl chloride; N-methylol compounds, e.g., N-methylol-chloro acetamide; di-isocyanates or di-isothiocyanates, e.g., phenylene-1,4-di-isocyanate and 30 aziridines. Other crosslinking agents include epoxides, such as, for example, di-epoxides, tri-epoxides and tetra-epoxides. For a representative listing of other available crosslinking reagents see, for example, the Pierce Catalog and Handbook, Pierce

- 20 -

Chemical Company, Rockford, Illinois (1997) and also S.S. Wong, Chemistry of Protein Conjugation and Cross-Linking, CRC Press, Boca Raton, Florida (1991).

According to one embodiment of this  
5 invention, carbohydrate crosslinked glycoprotein  
crystals are produced by a method comprising at least  
one crosslinking reaction through one or more  
carbohydrate moieties in said glycoprotein, alone or in  
sequence with crosslinking via a multifunctional reagent  
10 such as glutaraldehyde or other crosslinkers that  
function by crosslinking one or more amino acid side  
chain functional groups in said glycoprotein.

According to an alternate embodiment of this  
invention, at least one non-carbohydrate crosslinking  
15 reaction may be carried out using reversible  
crosslinkers, in parallel or in sequence with the  
carbohydrate crosslinking reaction. The resulting  
carbohydrate crosslinked glycoprotein crystals are  
characterized by containing a reactive multifunctional  
20 linker, into which a trigger has been incorporated as a  
separate group. The reactive functionality is involved  
in linking together reactive amino acid side chains in  
a glycoprotein and the trigger consists of a bond that  
can be broken by altering one or more conditions in the  
25 surrounding environment (e.g., pH, temperature, or  
thermodynamic water activity).

Examples of reversible crosslinkers are  
described in T.W. Green, Protective Groups in Organic  
Synthesis, John Wiley & Sons (Eds.) (1981). Any  
30 variety of strategies used for reversible protecting  
groups can be incorporated into a crosslinker suitable  
for at least one crosslinking in producing carbohydrate  
crosslinked glycoprotein crystals capable of

- 21 -

reversible, controlled solubilization. Various approaches are listed, in Waldmann's review of this subject, in Angewante Chemie Int. Ed. Engl., 35, p. 2056 (1996).

5 Other types of reversible crosslinkers are disulfide bond-containing crosslinkers. The trigger breaking crosslinks formed by such crosslinkers is the addition of reducing agent, such as cysteine, to the environment of the crosslinked protein crystals.

10 Disulfide crosslinkers are described in the Pierce Catalog and Handbook (1997). Examples of such crosslinkers include the symmetric homo-bifunctional, as for example DSS -dithiobis (succinimidyl-propionate), also known as Lomant's Reagent and DTSSP -  
15 3-3'-dithiobis (sulfo-succinimidylpropionate), a water soluble version of DSP and many more. Other examples include the heterobifunctional or asymmetric crosslinkers such as SPDP - N-succinimidyl-3-(2-pyridyldithio)propionate and LC-SPDP - succinimidyl-6-  
20 (3-[2-pyridyldithio] propionate)hexanoate and others.

In another embodiment of this invention, the dissolution, catalytic, antigenic and pharmaceutical properties of carbohydrate crosslinked glycoprotein crystals are modified by the specific combinations of  
25 crosslinking reactions performed.

Controlled dissolution of carbohydrate crosslinked glycoprotein crystals according to the present invention may also be effected by a change in time sufficient to permit a protein activity release rate between about 0.1% per day and about 100% per day, a change in time sufficient to permit a protein activity release rate between about 0.01% per hour and about 100% per hour or a change in time sufficient to

- 22 -

permit a protein activity release rate between about 1% per minute and about 50% per minute.

Carbohydrate crosslinked glycoprotein crystals according to this invention, therefore,  
5 include those capable of releasing their protein activity at a controlled rate upon exposure to a change in their environment, said change being selected from the group consisting of change in pH, change in solute concentration, change in temperature, change in  
10 chemical composition, change in shear force acting upon the crystals and combinations thereof.

Factors contributing to the release rate of protein activity of carbohydrate crosslinked glycoprotein crystals according to this invention  
15 include the degree of crosslinking of the carbohydrate crosslinked glycoprotein crystals, whether the reduction step was performed after the crosslinking reaction, the pH used for the oxidation reaction, the pH used for the crosslinking reaction, the pH used for  
20 the reduction reaction, whether the multifunctional crosslinking agent was pretreated, the polymerization state of the crosslinking agent, the number of crosslinking reactions performed, the length of time of exposure of protein crystals to the crosslinking agent,  
25 the rate of addition of crosslinking agent or agents to the glycoprotein crystals, the length of time of exposure of glycoprotein crystals to the crosslinking agent, the nature of the crosslinker either carbohydrate specific or amino acid side-chain  
30 specific, the chain length of the crosslinker, the surface area of the carbohydrate crosslinked glycoprotein crystals, the size of the carbohydrate crosslinked glycoprotein crystals, the shape of the

- 23 -

carbohydrate crosslinked glycoprotein crystals and combinations thereof.

According to this invention, any individual, including humans and other mammals, as well as birds 5 and fish, for example, may be treated in a pharmaceutically acceptable manner with a pharmaceutically effective or a catalytically effective amount of carbohydrate crosslinked glycoprotein crystals for a period of time sufficient to treat a 10 condition in the individual to whom they are administered over some period of time. Alternatively, individuals may receive a prophylactically effective or a catalytically effective amount of carbohydrate 15 crosslinked glycoprotein crystals which is effective to prevent a condition in the individual to whom they are administered over some period of time.

Carbohydrate crosslinked glycoprotein crystals may be administered alone, as part of a pharmaceutical, personal care or veterinary preparation 20 or as part of a prophylactic preparation, such as a vaccine, with or without adjuvant. They may be administered by parenteral or non-parenteral route. For example, they may be administered by oral, pulmonary, nasal, aural, anal, dermal, ocular, 25 intravenous, intramuscular, intraarterial, intraperitoneal, mucosal, sublingual, subcutaneous, or intracranial route. In either pharmaceutical, personal care or veterinary applications, carbohydrate crosslinked glycoprotein crystals may be topically 30 administered to any epithelial surface. Such epithelial surfaces include oral, ocular, aural, anal and nasal surfaces, to treat, protect, repair or detoxify the area to which they are applied.

- 24 -

Pharmaceutical combinations of the present invention may be formulated in a variety of conventional forms employed for parenteral administration. These include, for example, semi-solid and liquid dosage forms, such as liquid solutions or suspensions, suppositories, douches, enemas, gels, creams, emulsions, lotions, slurries, powders, and pastes. Standard formulation strategies for vaccines, immunotherapeutics and pharmaceuticals may be applied to carbohydrate crosslinked glycoprotein crystals in order to enhance the persistence and residence time of the active agent, and to improve the prophylactic efficacy achieved.

The present invention also includes controlled release formulations comprising carbohydrate crosslinked glycoprotein crystals. In such formulations, the carbohydrate crosslinked glycoprotein crystals are substantially insoluble under storage conditions and capable of releasing their protein activity in vivo at a controlled rate. For example, a pharmaceutical controlled release formulation according to this invention, administered by oral route, is characterized in that the component carbohydrate crosslinked glycoprotein crystals are substantially insoluble under gastric pH conditions and substantially soluble under small intestine pH conditions. Alternatively, for these and other uses according to this invention, the carbohydrate crosslinked glycoprotein crystals are biodegradable and may be active in the insoluble form and then dissolve and are removed or digested once their function is complete.

Pharmaceutical, personal care, veterinary or prophylactic compositions comprising carbohydrate crosslinked glycoprotein crystals according to this

- 25 -

invention may also be selected from the group consisting of tablets, liposomes, granules, spheres, microparticles, microspheres and capsules.

For such uses, as well as other uses  
5 according to this invention, carbohydrate crosslinked glycoprotein crystals may be formulated into tablets. Such tablets constitute a liquid-free, dust-free form of carbohydrate crosslinked glycoprotein crystal storage, which are easily handled and retain acceptable  
10 levels of activity.

Alternatively, the carbohydrate crosslinked glycoprotein crystals may be in a variety of conventional depot forms employed for administration to provide reactive compositions. These include, for  
15 example, solid, semi-solid and liquid dosage forms, such as liquid solutions or suspensions, gels, creams, balms, emulsions, lotions, slurries, powders, sprays, foams, pastes, ointments, salves, balms and drops.

According to one embodiment of this  
20 invention, carbohydrate crosslinked glycoprotein crystals may be combined with any conventional materials used for controlled release administration of pharmaceutical glycoproteins. Such materials include, for example, coatings, shells and films, such as  
25 enteric coatings and polymer coatings and films.

The most effective mode of administration and dosage regimen of formulations or compositions comprising carbohydrate crosslinked glycoprotein crystals of this invention will depend on the effect  
30 desired, previous therapy, if any, the individual's health status or status of the condition itself and response to the carbohydrate crosslinked glycoprotein crystals and the judgment of the treating physician or

- 26 -

clinician. The carbohydrate crosslinked glycoprotein crystals may be administered in any dosage form acceptable for pharmaceuticals, personal care compositions or veterinary formulations, at one time or 5 over a series of treatments.

The amount of the carbohydrate crosslinked glycoprotein crystals that may be combined with carrier materials to produce a single dosage form will vary depending upon the particular mode of administration, 10 formulation, dose level or dose frequency. A typical preparation will contain between about 0.01% and about 99%, preferably between about 1% and about 50%, carbohydrate crosslinked glycoprotein crystals (w/w).

Upon improvement of the individual's 15 condition, a maintenance dose of carbohydrate crosslinked glycoprotein crystals may be administered, if necessary. Subsequently, the dosage or frequency of administration, or both, may be reduced as a function of the symptoms, to a level at which the improved 20 condition is retained. When the condition has been alleviated, treatment should cease. Individuals may, however, require intermittent treatment on a long-term basis upon any recurrence of the condition or symptoms thereof.

25 The properties of the carbohydrate crosslinked glycoprotein crystals of this invention are especially suited for applications involving vaccines, to protect humans and animals from infectious diseases.

The methods of this invention may be used to 30 produce carbohydrate crosslinked glycoprotein crystals from virus, or parasite glycoproteins for use as antigens in a vaccine. Appropriately spaced and repeated parenteral inoculations of carbohydrate crosslinked glycoprotein crystals in combination with

- 27 -

immune system modifiers such as adjuvants and/or cytokines are useful to induce patient antibody and T-cell immune responses to viral, or parasite glycoproteins. The patient immune responses can 5 prevent or mitigate transmission or disease from of an enveloped virus infection, or parasite infections.

Such enveloped viruses include, for example, virus from the genera *Retroviridae*, *Herpesviridae*, *Orthomyxoviridae*, *Paramyxoviridae*, *Hepadnaviridae*, 10 *Flaviviridae*, *Togaviridae*, *Rhabdoviridae*, *Poxviridae*, *Arenaviridae*, *Coronoviridae*, *Bunyaviridae* and *Filoviridae*.

*Retroviridae* infections that may be prevented or treated using the methods of this invention include, 15 for example, those caused by human immunodeficiency virus type 1 and the human immunodeficiency virus type 2 lentiviruses, foamy viruses and human T-cell leukemia viruses.

*Herpesviridae* infections that may be 20 prevented or treated using the methods of this invention include, for example, those caused by herpes simplex type 1 and herpes simplex type 2 viruses, human herpes virus-6, Human herpes virus-8, varicella-zoster viruses, cytomegaloviruses, lymphoproliferative 25 herpesviruses, Epstein-Barr virus and other herpes viruses.

*Orthomyxoviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by 30 influenza A, influenza B and influenza C viruses.

*Paramyxoviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by

- 28 -

respiratory syncytial virus, mumps virus, parainfluenza viruses and measles-like viruses.

5 *Hepadnaviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by hepatitis B viruses.

10 *Flaviviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by hepatitis C virus, yellow fever virus, dengue virus and tick-borne encephalitis viruses.

15 *Togaviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by rubella virus.

20 *Rhabdoviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by rabies virus and vesicular stomatitis virus.

25 *Poxviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by vertebrate and avian poxviruses and vaccinia viruses.

30 *Arenaviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by the arenaviruses.

*Coronaviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by the coronaviruses.

35 *Bunyaviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by the hantaviruses.

- 29 -

*Filoviridae* infections that may be prevented or treated using the methods of this invention include, for example, those caused by Marburg, Reston and Ebola viruses.

5 Preferably, the enveloped viral infection to be treated or prevented is one wherein the virus is from the genera *Retroviridae*, *Herpesviridae*, *Orthomyxoviridae*, *Paramyxoviridae*, *Hepadnaviridae*, *Flaviviridae*, or *Rhabdoviridae*. More preferably, the 10 virus is type 1 or type 2 *human immunodeficiency virus*, type 1 or type 2 *herpes simplex virus*, *varicella zoster virus*, *Epstein-Barr virus*, *cytomegalovirus*, *influenza type A, B, or C virus*, *respiratory syncytial virus*, *mumps virus*, *hepatitis B virus*, *hepatitis C virus*, 15 *encephalitis virus*, *rabies virus*, or *dengue fever-inducing virus*. Most preferably, the virus is type 1 or type 2 *human immunodeficiency virus* or type 1 or type 2 *herpes simplex virus*.

20 Important examples of parasitic targets for which carbohydrate crosslinked glycoprotein crystal based vaccines include those in the kingdom *Protozoa*.

Protozoa infections that may be prevented or treated using the methods of this invention include those caused by representatives of the 25 *Sarcomastigophora* (containing flagellates and amebas); *Apicomplexa* (containing the sporozoans) and *Ciliophora* (containing the ciliates).

Sarcomastigophora infections that may be prevented or treated using the methods of this 30 invention include, for example, those caused by *Trypanosoma cruzi*, *Toxoplasma gondii*, *Leishmania major*.

- 30 -

Apicomplexa infections that may be prevented or treated using the methods of this invention include, for example, those caused by *Plasmodium falciparum*.

Ciliophora infections that may be prevented 5 or treated using the methods of this invention include, for example, those caused by *Balantidium coli*.

Parasitic worm infections for which vaccines may be effective include, for example, those caused by the classes *Cestoda* (tapeworms) and *Trematoda* (flukes).

10 According to one embodiment of this invention, the use of dried carbohydrate crosslinked glycoprotein crystals permits routine handling and storage of these materials prior to use (dry storage at room temperature or above without refrigeration, for 15 extended periods of time). The ability to transport the carbohydrate crosslinked glycoprotein crystals at ambient temperatures without denaturation of the glycoprotein advantageously overcomes the problem of inadequate refrigeration of vaccines often encountered 20 during global distribution of vaccines.

Dried carbohydrate crosslinked glycoprotein crystals also allow for routine formulation by direct addition of adjuvants and immune modifying cytokines such as Type I interferon, immune interferon, tumor 25 necrosis factor, interleukin-1, interleukin-2, interleukin-3, interleukin-4, interleukin-5, interleukin-6, interleukin-8, interleukin-10, interleukin-12, colony stimulating factors and other immune modifiers.

30 Any conventional pharmaceutically acceptable carrier or adjuvant may be combined with the carbohydrate crosslinked glycoprotein crystals of this invention. These carriers and adjuvants include, for

- 31 -

example, Freund's complete and incomplete, bacterial lipopolysaccharides, cholera toxin, mono and di-phosphoryl lipid A, ion exchangers, alumina, aluminum stearate, lecithin, serum proteins, such as human serum albumin,  
5 buffer substances, such as phosphates, glycine, sorbic acid, potassium sorbate, partial glyceride mixtures of saturated vegetable fatty acids, water, salts or electrolytes such as protamine sulfate, disodium hydrogen phosphate, sodium chloride, zinc salts,  
10 colloidal silica, magnesium, trisilicate, polyvinyl pyrrolidone, cellulose-based substances and polyethylene glycol. Adjuvants for topical or gel base forms may be selected from the group consisting of sodium carboxymethylcellulose, polyacrylates,  
15 polyoxyethylene-polyoxpropylene-block polymers, polyethylene glycol, natural and synthetic gum bases, and wood wax alcohols.

Formulations may include any excipient or carrier which may be added to carbohydrate crosslinked  
20 glycoprotein crystals or pharmaceutical salts thereof, without affecting their biological activity.

According to another embodiment, this invention provides a method for treating or preventing an enveloped virus infection in a patient comprising  
25 the step of administering to said patient a composition comprising: an amount of a carbohydrate crosslinked glycoprotein that is also an immune response modifier or antibody, sufficient to reduce or prevent viral replication in said patient via modification of  
30 patients immune response or direct interaction with said virus.

A carbohydrate crosslinked glycoprotein crystal or a combination of carbohydrate crosslinked glycoprotein crystals can be used as a component of a

sensor, referred to as a biosensor, useful for detecting and/or measuring an analyte of interest in a fluid, such as body fluid (e.g., blood, urine), chemical and laboratory reaction media, organic media, 5 water, culture medium and beverages. In some instances, the fluid in question can be a gas, as in an alcohol breath analyzer [E. Barzana, A. Klibanov and M. Karell, NASA Tech Briefs, 13, p. 104, (1989)]. In this application an appropriately-selected carbohydrate 10 crosslinked glycoprotein crystal is brought into contact with a fluid to be analyzed for the analyte of interest. The analyte of interest can be measured directly (e.g., blood glucose) or indirectly (e.g., by detecting or measuring a substance which is a reactant 15 (product or substrate) in a reaction in which the analyte of interest participates). In either case, the carbohydrate crosslinked glycoprotein crystal is able to act upon the analyte or the substance which is a reactant in a reaction in which the analyte also 20 participates. The activity of the enzyme results in a detectable change (e.g., change in pH, production of light, heat, change in electrical potential) which is detected and/or quantified by an appropriate detecting means (e.g., pH electrode, light or heat sensing 25 device, means for measuring electrical change) [J. Janette, et al., Anal. Chem., 62, pp. 33R-44R (1990)]. Any means useful for detecting the change resulting from the enzyme-catalyzed method can be used. A biosensor of the present invention includes a 30 carbohydrate crosslinked glycoprotein crystal or a combination of carbohydrate crosslinked glycoprotein crystals and a retaining means for the carbohydrate crosslinked glycoprotein crystal which allows contact between the carbohydrate crosslinked glycoprotein

- 33 -

crystal(s) and the analyte of interest or the substance in the fluid which is a reactant in the reaction in which the analyte of interest participates.

The carbohydrate crosslinked glycoprotein 5 crystals of this invention may be used in any of a number of chemical processes. Such processes include industrial and research-scale processes, such as organic synthesis of specialty chemicals and pharmaceuticals. Enzymatic conversion processes 10 include oxidations, reductions, additions, including esterifications and transesterifications, hydrolyses, eliminations, rearrangements, and asymmetric conversions, including stereoselective, stereospecific and regioselective reactions.

15 Carbohydrate crosslinked glycoprotein crystals according to this invention may also be used in various environmental applications. They may be used in place of conventional soluble or immobilized proteins for environmental purposes, such wide area 20 decontamination of environmental hazards.

Alternatively, the carbohydrate crosslinked glycoprotein crystals of this invention may be used in cleaning agents, selected from the group consisting of detergents, such as powdered detergents and liquid 25 detergents, bleaches, household cleaners, hard surface cleaners, industrial cleaners and carpet and upholstery shampoos.

Cleaning agents containing carbohydrate 30 crosslinked glycoprotein crystals according to the present invention may also comprise compounds conventionally included in such agents. See, for example, Soaps and Detergents. A Theoretical and Practical Review, Louis Spitz (Ed.), AOCS Press (Champlain, Illinois) (1996). Such compounds include

- 34 -

anionic, non-ionic, cationic or zwitterionic surfactants, or mixtures thereof.

Anionic surfactants are exemplified by alkyl sulfates, alkyl ether sulfates, alkyl sulfonates, 5 alkylaryl sulfonates, olefin sulfonates, alkyl ether phosphates, alkyl ether phosphates, fatty acid salts, soaps, isothionates and sulfonated unsaturated esters and acids.

Non-ionic surfactants are exemplified by 10 products of condensation of an organic aliphatic or alkyl aromatic hydrophobic compound with an alkylene oxide, alkyl polyglucosides and sugar esters.

Cationic surfactants are exemplified by 15 quarternary ammonium salts of tertiary alkyl amines, amino amides, amino esters or imidazolines containing at least one long chain ( $C_8-C_{22}$ ) aliphatic group or an alkyl-aryl group, wherein alkyl comprises about 4 to 12 carbon atoms and aryl is preferably a phenylene group.

Zwitterionic surfactants are exemplified by 20 derivatives of quarternary ammonium, quarternary phosphonium or tertiary sulfonium compounds, derivatives of secondary and tertiary amines and derivatives of heterocyclic secondary and tertiary amines.

Carbohydrate crosslinked glycoprotein 25 crystals according to this invention may also be used as ingredients in personal care compositions, including cosmetics, such as creams, lotions, emulsions, foams, washes, compacts, gels, mousses, slurries, powders, sprays, pastes, ointments, salves, balms, drops, shampoos and sunscreens. In topical creams and 30 lotions, for example, they may be used as humectants or for skin protection, softening, bleaching, cleaning, deproteinization, lipid removal, moisturizing,

- 35 -

decoloration, coloration or detoxification. They may also be used as anti-oxidants in cosmetics.

An alternate embodiment of the present invention includes protein delivery systems comprising 5 carbohydrate crosslinked glycoprotein crystals. Such a system may be used to deliver glycoproteins such as those included in personal care products, such as cosmetics, pharmaceuticals, veterinary compositions, vaccines, foods, feeds, diagnostics, cleaning agents, 10 such as detergents, and formulations for decontamination. Glycoprotein delivery systems of this invention, which may be formulations or devices, such as implantable devices, may be microparticulate glycoprotein delivery systems.

15 In such systems, as well as in other embodiments of the present invention, carbohydrate crosslinked glycoprotein crystals have a longest dimension between about 0.01  $\mu\text{m}$  and about 500  $\mu\text{m}$ , alternatively between about 0.1  $\mu\text{m}$  and about 50  $\mu\text{m}$ .

20 The crosslinked glycoprotein crystal components may have a shape selected from the group consisting of: spheres, needles, rods, plates, such as hexagons and squares, rhomboids, cubes, bipyramids and prisms.

Advantageously, the crosslinked crystal form of the 25 glycoproteins of this invention allow loading of up to between about 50% and about 90% protein per unit of weight.

According to one embodiment of this invention, carbohydrate crosslinked glycoprotein 30 crystals are characterized by stability and integrity under elution conditions used in separations, particularly chromatography elution conditions, as compared with the soluble uncrosslinked form of the protein that is crystallized to form the glycoprotein

- 36 -

crystals that are crosslinked. Alternatively, carbohydrate crosslinked glycoprotein crystals are characterized by stability and integrity in the presence of a solvent contained in the sample to be 5 separated, as compared with the soluble uncrosslinked form of the glycoprotein that is crystallized to form the glycoprotein crystals that are crosslinked.

The carbohydrate crosslinked glycoprotein crystals may be used for high throughput screening in 10 combinatorial chemistry, where large libraries may be screened for specific interaction with the glycoprotein component of the crystals.

An advantage of this invention is that chromatographic separations may be carried out in the 15 presence of an aqueous solvent, an organic solvent, or an aqueous-organic solvent mixture.

In order that this invention may be better understood, the following examples are set forth. These examples are for the purpose of illustration only 20 and are not to be construed as limiting the scope of the invention in any manner.

#### EXAMPLES

##### Example 1 - Preparation of Glucose Oxidase Crystals

We prepared crystals of glucose oxidase as 25 follows. First, the glycoprotein was purified by anion exchange chromatography and then the crystallization parameters were optimized.

###### A. Preparative chromatography of glucose oxidase

One volume of 50 ml of crude glucose oxidase 30 "GO" (Novo Nordisk Bioindustrials, Franklinton, North Carolina) was diluted with 4 volumes, 200 ml, of 25 mM

- 37 -

Na-acetate at a pH of 4.5. The solution was diafiltered with Fresenius F 40 hollow fiber ultrafilter to equilibrium with the buffer.

Glycoprotein concentration was expressed as ultraviolet 5 absorbance at wavelength 280 nm, and abbreviated as A 280 nm. The A 280 nm of a sample is reproducible and is directly proportional to the glucose oxidase concentration when the enzyme sample is pure, i.e. contains only glucose oxidase glycoprotein. After 10 diafiltration, the volume was 265 ml with an A 280 nm equal to 18.9. Then 160 ml of the filtered solution was slowly pumped onto a Pharmacia XK 16 chromatography column with a 16 mm inner diameter, 20 cm length and 14 cm bed height packed with 28 ml of Q-sepharose. The 15 column was washed with 58 ml of the 25 mM Na-Acetate pH 4.5 running buffer prior to elution. Glucose oxidase was eluted with 80 mM NaCl and stored at 4°C for use in crystallization experiments. Glucose oxidase purchased from Sigma Chemical Co., St. Louis, Mo, was also 20 purified for use in crystallization experiments.

#### B. Optimization of Crystal Formation

##### 1.0 Effect of polyethylene glycol molecular size

We analyzed the effect of polyethylene glycol "PEG" molecular size on crystal formation in small 25 scale microdiffusion experiments using the Hanging Drop method. In the Hanging Drop method, the drop hangs with surface tension on a cover slide over a chamber filled with crystallizing media. The drop consists of 30 5 µl of enzyme solution sample plus 5 µl of the chamber solution. The chamber is sealed tightly with the cover slide (hanging drop inside). Diffusion concentrates the drop slowly to a volume of 5 µl and allows

- 38 -

supersaturation of the glycoprotein. The volume pipetted into the chamber is typically 400-500  $\mu$ l.

The following molecular sizes were evaluated; PEG 400, 600, 1000, 4000, 6000, or 8000 at four 5 different concentrations each. The PEG 400 varied from 20-50% final concentration. The PEG 600 varied from 9-20% concentration. The PEG 1000 varied from 8-15% final concentration. The PEG 4000 varied from 7-13% final concentration. The PEG 6000 varied from 6-10% 10 final concentration. The PEG 8000 varied from 4-8% final concentration. The pH was kept constant at 5.0 and citric acid concentration varied from 0.1 M to 0.2 M. The initial glycoprotein concentration was A<sub>280</sub> of 14.4 and increased to A<sub>280</sub> of 28.8, while the lithium 15 chloride (LiCl) concentration varied from 1 to 2 Molar. We previously determined that an A<sub>280 nm</sub> = 1.0 for glucose oxidase indicates that the concentration of glucose oxidase is 0.714 mg/ml.

Large crystals of glucose oxidase were 20 produced with PEG 600, 1000, 4000, and 6000. The amorphous precipitate form was favored with the high LiCl concentration, making complete crystallization difficult. We next evaluated the effects of LiCl and pH on crystal formation.

25           2.0 Effect of pH and LiCl

The pH was then varied between 5.0, 5.5 and 6.0, to determine the optimum pH for glycoprotein crystallization. Crystallization was carried out within this range in either the presence or absence of 30 1.0 M LiCl, that was added gradually by hanging drop. The maximum protein concentration reached an A<sub>280</sub> of 24.2 or 24.5 and the PEG 6000 varied from 10-20% final concentration. The entire experiment was carried out

- 39 -

with both Sigma Chemical Co. [SI] and Novo Nordisk [NO] glucose oxidase from Aspergillus niger. Crystals could be obtained without LiCl, in solutions containing between 10 and 20% PEG.

5        3.0 Effect of PEG size without LiCl

We next evaluated the effect of removing LiCl on crystal formation using four different molecular sizes of PEG: PEG 200, 400, 600, 1000, 4000, or 6000, at four concentrations. The PEG 200 varied from 15-30% final concentration. The PEG 400 varied from 10-25% final concentration. The PEG 600 varied from 8-20% final concentration. The PEG 1000 varied from 5-15% final concentration. The PEG 4000 varied from 5-13% final concentration. The PEG 6000 varied from 5-10% final concentration. The pH was kept constant at 5.0 and the Na-citrate buffer was 0.1 M. The initial glycoprotein concentration was A280 of 13.1 and increased to A280 of 26.2.

As a result of the small scale hanging drop microdiffusion studies, we determined that optimal crystals were obtained using either 20% PEG 4000 or PEG 6000, in the absence of LiCl.

4.0 Batch Crystallization

We carried out batch crystallizations of glucose oxidase using reagents prepared at their final concentrations and incubation under the specified conditions. Small batches having different PEG 4000 concentrations were prepared in 1.5 ml Eppendorf tubes. The samples were buffered in 0.1 M Na-Acetate at pH 5.0 and rocked at room temperature. The results are shown in Table 1 below, in which "C" designates crystals, C\* indicates the most uniform crystals obtained and "L" designates liquid phase separation.

- 40 -

**Table 1**

PEG %	50% PEG 4000 in buffer	Enzyme SI in buffer μl	Initial A280	Soluble A280 after 3 days	results after 3 days
5	15	300	700	27.7	18.4
	16	320	680	26.9	10.1
	17	340	660	26.1	5.2
	18	360	640	25.3	3.3
	19	380	620	24.6	2.0
	20	400	600	23.8	1.6

10 Crystallization conditions in the samples having 16% or higher PEG concentration produced very good crystals.

#### 5.0 Addition of Organic Solvents

15 The addition of water miscible organic solvent to a solution containing glycoprotein, PEG and buffer reduces the viscosity and the amorphous gel precipitation. Therefore, concentrations of PEG were chosen at the low end of the effective crystallization range. Both 15% and 17% PEG 6000 and, in a few cases, 20 PEG 4000 were used in batch crystallizations at room temperature.

A stock solution of 1.5 g of glucose oxidase in 50 ml of 0.2 M Na-acetate at pH 5.0 was prepared. The A280 of this solution was 46. The crystallizations 25 were begun by mixing 0.5 ml of protein with 0.5 ml of the PEG-solvent mixture, which had twice the final concentration indicated. The final PEG concentration was 15% in each experiment. The solvents were used at final concentrations of 5, 10 or 15%. The results are

- 41 -

shown in Tables 2 and 3 below, in which "MPD" designates 2-methyl-2,4-pentanediol, "A" designates amorphous, "C" designates crystals, "Cg" designates granule-like crystals and "L" designates liquid phase separation.

**Table 2**

Effect of Solvents in 15% PEG 6000 at Room Temperature

Constant parameters				
Sample SI, A 280 = 23 Temp = RT Precipitant = 15% (V/V) PEG 6000				
Exp	Solvent	Conc (V/V)	Supernat A280 at 6 days	Microscopy Results after 5 days
10	1 Ethanol	5%	5.05	G
	2 Ethanol	10%	1.84	C
	3 Ethanol	15%	0.95	C
	4 2-Propanol	5%	0.87	C
	5 2-Propanol	10%	2.25	Cg
15	6 2-Propanol	15%	4.64	G
	7 Acetone	5%	*	Cg
	8 Acetone	10%	*	Cg
	9 Acetone	15%	*	Cg
	10 MPD	5%	10.07	Cg
20	11 MPD	10%	11.82	Cg
	12 MPD	15%	12.19	Cg
	13 Ethylene glycol	5%	5.37	Cg
	14 Ethylene glycol	10%	2.69	N
	15 Ethylene glycol	15%	1.68	C
25	16 Water		9.16	Cg

30 Water miscible organic solvents greatly improved the crystallization, particularly ethanol and 2-propanol. The other combinations produced only granular sticky gel precipitates. The reference sample with PEG alone did not yield any crystals.

- 42 -

**Table 3**

Effect of Solvents in 17% PEG 6000 at Room Temperature

Constant parameters				
Sample SI GO				
A 280 = 23 Temp = RT				
Precipitant = 17% (V/V) PEG 6000				
Exp.	Solvent	Conc (V/V)	Supernat A280 at 6 days	Microscopy Results after 5 days
17	EtOH	5%	1.58	C
18	EtOH	10%	0.74	C
19	EtOH	15%	0.67	C
20	2-Propanol	5%	1.54	C
21	2-Propanol	10%	1.05	C
22	2-Propanol	15%	0.72	Cg
23	Acetone	5%	*	C
24	Acetone	10%	*	C
25	Acetone	15%	*	C
26	MPD	5%	2.01	N
27	MPD	10%	1.25	N
28	MPD	15%	1.11	Cg
29	Ethylene glycol	5%	4.31	Cg
30	Ethylene glycol	10%	5.21	Cg
31	Ethylene glycol	15%	5.8	Cg
32	Water		4.13	Cg

This series of examples produced more complete crystallization than the previous set (Table 2). Three of the solvents induced good crystal formation namely, ethanol, 2-propanol, and acetone. On microscopic analysis, we determined that the crystals were better and the crystallization process more complete than in the previous set of examples.

- 43 -

#### 6.0 The Effect of Solvents at 4°C

The samples were mixed as in the previous example using 15% PEG 6000 and rocked in the refrigerator at 4°C. The results are shown in Table 4  
5 below, in which "C" designates crystals, and "Cg"  
designates granule like crystals.

**Table 4**

#### Effect of Solvents in 15% PEG 6000 at 4°C

		Constant parameters		Sample = SI GO	
		A 280 = 23, Temp = 4 °C		Precipitant = 15% (V/V) PEG 6000	
	Exp	Solvent	Conc (V/V)	Supernat A280 at 6 days	Microscopy Results after 5 days
10	33	EtoH	5%	3.27	CgC
	34	EtoH	10%	1.56	CgC
	35	EtoH	15%	1.1	C
	36	2-Propanol	5%	0.71	Cg
	37	2-Propanol	10%	1.65	C
	38	2-Propanol	15%	2.54	CgC
15	39	Acetone	5%	*	C
	40	Acetone	10%	*	C
	41	Acetone	15%	*	Cg
	42	MPD	5%	4	Cg
	43	MPD	10%	2.28	Cg
	44	MPD	15%	1.05	Cg
20	45	Ethylene glycol	5%	7.3	Cg
	46	Ethylene glycol	10%	7.49	Cg
	47	Ethylene glycol	15%	8.09	Cg
	48	Water		6.2	Cg

The reactions containing ethanol, 2-propanol  
30 and acetone all produced good crystals. The result was  
greatly improved over that obtained by incubation at  
room temperature.

The crystallization shown below in Table 5  
was performed as described for Table 3, except that the

- 44 -

samples were incubated and rocked at 4°C. In Table 5, "C" designates crystals, and "Cg" designates crystal-like granules.

**Table 8**

**5 Effect of Solvents in 17% PEG 6000 at 4°C**

		Constant parameters      Sample = SI GO A 280 = 23, Temp = 4 °C Precipitant = 17% (V/V) PEG 6000		
		Conc (V/V)	Supernat A280 at 6 days	Microscopy Results after 5 days
Exp	Solvent			
10	49	EtoH	5%	1.48
	50	EtoH	10%	1
	51	EtoH	15%	0.8
	52	2-Propanol	5%	1.27
15	53	2-Propanol	10%	0.88
	54	2-Propanol	15%	0.75
	55	Acetone	5%	*
	56	Acetone	10%	*
20	57	Acetone	15%	*
	58	MPD	5%	1.39
	59	MPD	10%	1.18
	60	MPD	15%	0.94
25	61	Ethylene glycol	5%	2.89
	62	Ethylene glycol	10%	3.07
	63	Ethylene glycol	15%	4.26
	64	Water		2.59

The samples containing the solvents ethanol, 2-propanol and acetone produced good crystals.

However, the results were not improved over that are obtained at room temperature. One difference was that 30 the sample with MPD produced crystals, a result not obtained at room temperature.

In summary, the addition of organic solvents resulted in dramatic improvements in crystal quality and quantity and also reduced the amorphous precipitate formation. Refrigeration seemed to improve the 35

- 45 -

situation even further. Ethanol and 2-propanol were selected for additional optimization.

7.0 Optimization of PEG 6000 and 2-Propanol at Reduced Protein Concentration

5 A stock solution of 50% PEG 6000 was prepared in 0.1 M Na-acetate at pH 5.0. The glucose oxidase stock solution was prepared with 1.5 g of Sigma Chemical Co. glycoprotein in 50 ml of 0.2 M Na-acetate. The A280 of the glucose oxidase solution was 46. 2-  
10 Propanol was used as 100 %. These solutions were mixed to produce the various final combinations: Crystals were observed only in solutions with 15% PEG and 10 or 15% 2-propanol. All other samples had amorphous precipitate or gel granules. The soluble protein level  
15 declined to less than 10% of the initial value in the samples with crystals, thus indicating that 90% or more protein was either crystallized or precipitated.

The above crystallization conditions were used with twice the input protein level or A280 of 20.  
20 Under these conditions, crystals were observed only in solutions having 15% PEG and either 5 or 10% 2-propanol. The other samples had only amorphous precipitate. Under the conditions that produced crystals, the A280 value of the supernatant indicates  
25 that 83-93% of the protein was in the form of crystals and precipitate.

The same conditions discussed above were employed with PEG 4000 instead of PEG 6000. The effect of PEG 4000 was evaluated with 2-propanol  
30 concentrations of 5, 10, and 15%. A stock solution of 50% PEG 4000 was prepared and the solutions were mixed to achieve the same conditions as discussed above.

- 46 -

This example was carried out at room temperature, in 0.1 M Na-acetate.

In the first example, 10 mg/ml glycoprotein was used. The samples having 15% PEG 4000 and 10 or 5 15% 2-propanol were crystallized. All other combinations produced only amorphous precipitate. The crystallized samples showed that 85-92% of protein converted into solid phase including crystals and precipitate. In general, the results were very similar 10 to those achieved using PEG 6000.

The next example used a glycoprotein content of equal to an A280 of 20 in the conditions discussed above. Only the sample having 15% PEG 4000 and 10% 2-propanol produced crystals. The other samples had only 15 amorphous precipitate. In the sample with crystals, 88% of the glycoprotein was in the solid phase.

In summary, PEG 4000 and PEG 6000 both are suitable for inducing glycoprotein crystal formation. The combination of 15% PEG 4000 or PEG 6000 and 10-15% 20 2-propanol, using relatively low protein levels, was remarkably effective in producing good quality glycoprotein crystals. By microscopy, all the crystallized samples had initially lumps of glassy gel precipitate prior to crystal growth. It appeared as 25 though the crystals were growing out of the surface of the gel particles.

#### 8.0 Optimization of PEG 6000 and Ethanol at Reduced Protein Concentration

The potential of ethanol in combination with 30 PEG 6000 as glycoprotein precipitants was evaluated. A total of 2.5 g of glucose oxidase glycoprotein from Sigma Chemical Co. was dissolved in 25 ml water and the solution was filtered through a 0.2  $\mu\text{m}$  membrane. This

- 47 -

enzyme stock solution had an A<sub>280</sub> of 139.4. A 50% PEG 6000 stock solution was prepared in 0.1 M Na-acetate at pH 5.0. Batch crystallization samples were prepared in Eppendorf tubes by mixing the following reagents:

- 5        1. 0.1 ml of glycoprotein stock solution (each batch final A<sub>280</sub> = 13.9),
2. sufficient PEG 6000 stock solution to reach the target concentration of 0 to 19%,
3. sufficient 100% ethanol to get to the target concentration of 0-21%; and
- 10      4. 0.1 M Na-acetate to reach the final volume of 1.0 ml.

The samples were rocked at room temperature and inspected by microscopy 1, 3 and 8 days after mixing.  
15 Finally, the samples were centrifuged at 10,000 RPM for 15 minutes and the soluble protein in supernatant was measured as A<sub>280</sub> nm.

Glycoprotein crystals were produced under conditions where the PEG 6000 concentration was 15 or  
20 17% and the ethanol concentration was 5-21%. The best ethanol conditions for glycoprotein crystallization were between 11-17%. PEG 6000 alone or ethanol alone did not induce crystal formation. Typical soluble protein levels in the samples that were well  
25 crystallized was around an A<sub>280</sub> of 1.0. Thus, in most of the crystallized samples greater than 90% of the protein was in the solid phase, crystals and precipitate. Under the best conditions, crystallization occurred overnight.

30      9.0 Preparative batch crystallizations with PEG 6000 and 2-propanol

Once the total volume of the batches increased from 1 ml to 5 ml, gel formation became

- 48 -

- somewhat problematic. The best conditions for crystal formation without gel formation were re-evaluated. Four different PEG 6000 concentrations were screened with 2-propanol from 3-20% (V/V) in 0.1 M Na-acetate.
- 5 The 5.0 ml batch samples were prepared and rocked at 6 °C, and evaluated by microscopy after days 1, 2, and 8. The results are shown below in Table 6, in which "A" designates amorphous, "C" designates crystals, "G" designates granule gel, "N" designates no phase separation and "L" designates liquid phase separation.
- 10

- 49 -

Table 6

Optimization of conditions for 5.0 ml  
preparative scale crystallizations

Exp	Precipitants		A 280 at 9 days	Microscopy Results After days				
	2-propanol Conc (V/V)	PEG 6000 Conc (W/V)		0	1	3	8	
5	1	10%	6%	18.3	N	C	-	C
	2	12%	6%	17.3	N	LC	-	C
	3	14%	6%	15.9	N	L	-	C
	4	16%	6%	14.4	N	L	-	C
	5	18%	6%	10.4	N	C	N	C
10	6	20%	6%	8.0	N	GC	C	CG
	7	8%	9%	12.9	N	C	-	C
	8	10%	9%	8.8	N	L	C	CG
	9	12%	9%	6.8	A	CG	C	C
	10	14%	9%	6.4	A	CG	C	CG
15	11	16%	9%	4.9	A	AC	C	C
	12	18%	9%	4.5	A	CG	C	C
	13	6%	10%	9.4	A	L	C	CG
	14	8%	10%	7.0	A	CG	C	CG
	15	10%	10%	5.9	A	CG	C	CG
20	16	12%	10%	5.4	A	CG	C	CG
	17	14%	10%	4.7	A	LCG	C	C
	18	3%	12%	6.8	A	LC	C	CG
	19	4%	12%	6.2	A	CG	C	C
	20	5%	12%	5.7	A	CG	C	C

As shown in Table 6, the combination of 9% PEG 6000 and 12% 2-propanol produced rapid crystal formation without gel.

#### Summary

As a result of these studies, we believe that the conditions for crystallization of glucose oxidase fall generally in the range of 7 to 17% PEG 4000 or 6000, 8 to 20% 2-propanol or ethanol, and buffer to adjust the pH to between 3 and 6. It should be understood, however, that many sets of experimental conditions within and near this range can produce satisfactory results for the crystallization of glucose

- 50 -

oxidase and other glycoproteins. Those of skill in the art will appreciate that the precise conditions which efficiently produce crystals of the desired size and quality, will vary due to differences in experimental 5 conditions, such as protein and reagent purity, rates of stirring, shear force effects, and carbohydrate content.

Example 2 - Large Scale Crystallization of Glucose Oxidase

10 Based on the optimization procedures outlined in Example 1, we determined preferred conditions for preparative scale crystallization of glucose oxidase. Preparative scale crystallization generally involves 100 to 900 mls of glycoprotein. The procedures are 15 exemplified below:

A. Crystallization at Constant pH Without Seed

Glucose oxidase was diafiltered in water and concentrated to an A 280 nm of between 5 and 15. The 20 glucose oxidase concentrate was mixed (1:1) with one volume of the crystallizing reagent containing 18% PEG 6000, 32% 2-propanol in 0.2 M Na-Acetate at pH 5.0. After mixing, the solution was cooled to 6 °C. The glucose oxidase crystallization solution was 25 stirred for 24 hours at 100 rpm with a propeller stirrer. During this time, the crystals formed gradually.

B. Crystallization at Constant pH With Seed Crystals

30 This method is similar to the "crystallization at constant pH without seed crystals"

- 51 -

method described above, except that seed crystals from previous batches were added after mixing the glycoprotein concentrate and crystallizing reagent. Crystallization occurred more rapidly by this method  
5 and there were many crystals after 1 hr of mixing.

C. Crystallization by Gradual pH Reduction

This method was carried out at a higher pH than the two examples shown above. The supersaturation of the solution was slowly increased by gradually  
10 lowering the pH to the point of lowest glucose oxidase solubility at pH 4.5.

The initial crystallization mixture included 35 ml of glucose oxidase stock solution with A 280 nm of 103, 337 ml water, 400 ml crystallizing reagent.  
15 The crystallizing reagent contained 180 g PEG 6000, 320 ml 2-propanol, 0.145 moles Na-Acetate and diluted to 1000 ml. The initial pH is usually between 6.5 and 7.0. The initial pH does not allow crystallization.

Crystallization was initiated by pumping 28.4  
20 ml of 1 M acetic acid into the batch with a flow rate of 2.2 ml/hr. The mixture was slowly cooled to 5 °C over 12 hours. The final pH was 4.5 and was stirred at 100 rpm.

Glucose oxidase crystallized during the first  
25 24 hours and formed crystals of between 10 to 15 µm. This method reproducibly produced the optimal and most uniform crystals.

Example 3 - Carbohydrate Crosslinking of Glucose Oxidase Crystals

30 We then crosslinked the glucose oxidase crystals prepared in Example 1 as follows. More particularly, we crosslinked the crystals by first

- 52 -

using oxidation of the carbohydrate, followed by crosslinking with a diamine compound and then finally reduction of the imine. The reduction step is one variable which can be used to control the relative  
5 insolubility or rate of dissolution of the carbohydrate crosslinked glycoprotein crystals. If controlled dissolution carbohydrate crosslinked glycoprotein crystals are the goal, then the reduction step can be left out. However, if insoluble carbohydrate  
10 crosslinked glycoprotein crystals are desired then the reduction step should be included. This series of reactions causes the carbohydrate moieties within and between protein molecules to become covalently crosslinked, thus stabilizing the crystalline form of  
15 the glycoprotein. According to an alternate embodiment of this invention, carbohydrate crosslinking may be combined with other carbohydrate crosslinking reactions or reactions that crosslink through amino acid side chain functional groups of the amino acid residues of  
20 the protein backbone. It should be understood that the particular combinations of these reactions determine the level of protein activity within the crystal, the rate of dissolution, the preservation of antigenic and immunogenic epitopes and the resistance to proteases,  
25 organic solvents, extremes of temperature and pH.

A. Crosslinking of Glucose oxidase through the attached carbohydrate

1.0 Oxidation of carbohydrates

Crystalline glucose oxidase, 102 mg, as  
30 prepared in Example 2, using method B was suspended in crystallization buffer consisting of 9% PEG 6000, 12% 2-propanol, 0.1 M Na-acetate at pH 5.0 at 6°C. Then 68 to 586 mg of sodium periodate ( $\text{NaIO}_4$ ) was added per

- 53 -

gram of glycoprotein crystal. All samples were protected from the light throughout the oxidation period. To carry out the oxidation, samples 1-4, shown in Table 7, were rocked for 2 hours at room 5 temperature, samples 5 and 6, were rocked overnight at 4°C. The crystals were centrifuged at 10,000 RPM for 10 minutes and then washed three times with crystallization buffer. Controlled dissolution activity of the crystals, shown in Table 7, was 10 measured by separating them by filtration after 30 minutes of titration and measuring the enzyme activity from the filtrate. In Table 7, the dissolved activity is presented as a percentage of the total activity after 30 minutes of titration.

15           2.0 Reaction with diamine

The crystals were dispersed in a diamine reagent solution consisting of 0.25 M 1,8-diaminoctane, 9% PEG 6000, 16% 2-propanol in 0.1 M Na-acetate at pH 5.0. The reaction was allowed to proceed 20 overnight at room temperature.

3.0 Reduction of imine compound

The amine reaction was terminated by addition of 760 mg of solid NaCNBH<sub>3</sub>, per gram of protein directly on the reaction mixture and incubated 90 minutes at 25 room temperature. Crystals were centrifuged and washed four times with 2 mM sodium phosphate, pH 7.0.

- 54 -

**Table 7**

Protein activity of carbohydrate  
crosslinked glycoprotein crystals

5	Sample No.	NaIO <sub>4</sub> , mg/g GO	Titrimetric activity U/ $\mu$ g		Activity of filtrate U/ $\mu$ g	Controlled dissolut. activity % of total
			Titr. 10 min	Titr. 30 min		
	1.	69	135	206	152	74
	2.	147	85	102	74	72
	3.	284	99	155	108	70
10	4.	588	114	164	98	60
	5.	147	180	271	-	-
	6.	441	123	131	88	67

**4.0 Measurement of glycoprotein activity**

The glycoprotein enzyme known as glucose oxidase, also known as  $\beta$ -D-glucose 1-oxidoreductase (enzyme commission number 1.1.3.4) catalyzes the oxidation of  $\beta$ -D-glucose to D-glucono-1,5-lactone and hydrogen peroxide ( $H_2O_2$ ). The reaction consumes one mole of oxygen per mole of glucose. The D-glucono-1,5-lactone subsequently hydrolyses to gluconic acid and the appearance of which in solution may be used to measure enzyme activity, as described below.

**4.1 Titration of protein activity**

The activity of the carbohydrate crosslinked glucose oxidase crystals was measured by titrating the production of gluconic acid. The analysis was performed in a 1000 ml reaction vessel which was immersed in 30°C temperature controlled water bath. The approximate diameter of the liquid was 95 mm and depth 60 mm. The batch was agitated with overhead propeller stirrer equipped with digital rpm indicator.

- 55 -

The stirrer was a 55 mm diameter 6 blade propeller. Air was dispersed in the batch through two pieces of 13 mm x 13 mm Pyrex No. 1 glass sinter beads. Air was pumped with Watson Marlow 603S peristaltic pump using 5 9.5 mm internal diameter pump hose. The process was controlled with Radiometer PHM 290 titrator. The results were collected with a computer which was connected to the titrator. The titrator control settings were the following:

10        Point                          1 min  
            Time constant                5.0 sec  
            Gain                            0.01  
            Set point                     pH 6.0, if not otherwise stated.

15        Specifics of the Procedure Include:

1.      Process volume was 400 ml in the start of titration.
2.      Glucose 100 g/l in deionized water, no buffer added
- 20     3.      carbohydrate crosslinked glucose oxidase crystals as dry substance in the batch
4.      Catalase 0.1 ml (500 units), Genencor CAT HP L5000
5.      Stirring 1000 rpm
- 25     6.      Aeration 2.4 l/min.
7.      Temperature 30°C
8.      The titration was made with 1 M or 10 M NaOH.
9.      Set point pH 6.0.

30      The total quantity of glucose was 0.222 moles in the standard analysis; thus the total consumption of 10 M NaOH was 22.2 ml, if the reaction was allowed to proceed to 100% conversion. The standard conditions were used if not otherwise stated.

- 56 -

This example demonstrates carbohydrate crosslinking of glycoproteins. The crystals subjected to the oxidation reaction with diamine and reduction reactions were all active and relatively insoluble 5 during extensive washing.

By virtue of their controlled dissolution protein activity, carbohydrate crosslinked crystals of glucose oxidase produced using this method of crosslinking would be preferred in the preparation of 10 crosslinked glycoproteins from viruses, parasites, tumors and cytokines, as well as other immune modifying glycoproteins for use in vaccines and immunotherapeutic applications.

A. Double Crosslinking of Glucose Oxidase Crystals

1.0 Carbohydrate crosslinking

According to an alternate embodiment of this invention, carbohydrate crosslinked glycoprotein crystals may be of minimal solubility. In order to 20 reduce the dissolution of protein activity from the single crosslinked glycoprotein crystals described above, the singly crosslinked crystals were submitted to a second crosslinking procedure. The second crosslinking procedure involved glutaraldehyde, or 25 glutaraldehyde pretreated with either Tris buffer (2-amino-2-hydroxymethyl-1,3-propanediol), lysine or diaminoctane.

The crosslinking parameters for the first carbohydrate crosslinking were the same as for sample 5 and 6 described above, and shown in Table 7. The 30 oxidation and reaction with diaminoctane steps were followed by rocking overnight at 4°C. In each case, the starting reaction contained 60 mg of protein.

- 57 -

**Table 8**

Protein activity of carbohydrate  
crosslinked glycoprotein crystals

No.	NaIO <sub>4</sub> mg/g GO	Titrimetric activity Glucose Oxidase 'U/ $\mu$ g			Total protein soluble in 0.1 M NaOH $A_{278}$	Solub. in NaOH %
		Titr. 10 min	Titr. 30 min	After control dissol. test		
5	1A.	36	299	494	195	1.8
	2A.	72	258	508	274	1.8
	3A.	143	195	453	253	1.9
	4A.	143	76	88	151	20.3
						33.8

The data presented in Table 8 indicate that oxidation with lower concentrations of NaIO<sub>4</sub> preserved and enhanced the protein activity of glucose oxidase in the resulting crystals most effectively. The titrimetric activities were measured after 30 minutes for crystals oxidized with either 36 or 72 mg/g GO and shown in Nos. 1A and 2A of Table 8, were 494 and 508 U/ $\mu$ g respectively. These values were almost twice the value of 279 U/ $\mu$ g obtained from GO crystals before crosslinking. The approximately 2-fold increase in glycoprotein activity obtained between sample 2A and the non-crosslinked material demonstrates that very significant enhancements of glycoprotein activity may be achieved through the carbohydrate crosslinking methods of this invention. Sample 3A was treated exactly as samples 1A and 2A except with more NaIO<sub>4</sub>. For sample 4A, the washing step after oxidation was eliminated and the diaminooctane was added directly to the overnight oxidation reaction mixture. Based on

- 58 -

this result we conclude that washing of the crystals after oxidation and before crosslinking is critical.

#### 1.1 Solubility in 0.1 M NaOH

The solubility of the crosslinked crystals in  
5 0.1 NaOH was measured using 1 ml of crosslinked  
crystals. The crystals were initially centrifuged and  
then resuspended in 1 ml of 0.1 M NaOH. After a two  
hour incubation, the supernatant was clarified by  
centrifugation and the protein A 278 was measured from  
10 supernatant, as shown in Tables 8 and 9.

#### 1.2 Solubility in Phosphate Buffer

We measured the solubility of the double  
crosslinked glycoprotein crystals using a controlled  
dissolution test. A 1 ml sample of crosslinked  
15 crystals was suspended in 2 mM sodium phosphate buffer  
(pH 7.0) and then diluted in 4 ml of 0.2 M glucose in 2  
mM sodium phosphate buffer at pH 7.0. The crystal  
samples were then incubated overnight at room  
temperature. Next the crystals were collected by  
20 centrifugation at 5,000 rpm for 10 min, after which  
activity was measured by titration and the results are  
shown in Tables 8 and 9. After the first crosslinking,  
approximately one-half of the glucose oxidase catalytic  
activity was dissolved in the substrate buffer during  
25 the controlled dissolution test which lasts 30 minutes.  
The other half remained with the crystals.

#### 2.0 Amino acid side chain functional group crosslinking

The second crosslinking was performed using  
30 aliquots of the most active sample (no. 2A.) of the  
first crosslinking experiment shown in Table 8.  
Glycoprotein amino acid side chain functional group  
crosslinking reagents were added to the carbohydrate

- 59 -

crosslinked glycoprotein crystals suspended in 0.2 M sodium phosphate at pH 7.0. Reactions were allowed to proceed for two hours at room temperature, with each second crosslinking reagent described below. We found  
5 that centrifugation was not a preferred method for recovering all the crystal material after washing. Therefore the crystals were filtered and washed over glass fiber paper.

- Protein amino acid second crosslinking  
10       conditions for samples shown in Table 9:
- 1B and 2B:     Diluted glutaraldehyde was added to final concentrations of 0.2 and 0.6 g/g GO, respectively.
- 15       3B and 4B:     Tris pretreated glutaraldehyde (48 mg Tris-base/g glutaraldehyde) was added to concentrations of 0.2 and 0.6 g/g GO crystals, respectively. In sample 4B., the washing steps after oxidation were eliminated and the diaminooctane was  
20       added directly to the oxidation mixture.
- 25       5B and 6B:     First lysine and then glutaraldehyde were added to molar ratio of 4:10. Final glutaraldehyde concentrations were 0. 2 and 0.6 g/g GO.
- 30       7B and 8B:     Same molar ratio of glutaraldehyde and diaminooctane were added by having 15 minute incubation time between each addition. Both were added two times. Total glutaraldehyde dosages were 0.2 and 0.6 g/g GO.

- 60 -

**Table 9**

Protein activity of carbohydrate and  
protein crosslinked glycoprotein crystals

	Sample No.	Titrimetric activity U/ $\mu$ g Glucose oxidase (GO)			Total protein soluble in 0.1 M NaOH A <sub>270</sub>	Solub. in NaOH %
		Titr. 10 min	Titr. 30 min	After Controlled Dissolut. test		
5	1B.	108	148	143	4.0	6.6
	2B.	110	187	156	3.9	6.6
	3B.	111	210	112	4.0	6.7
	4B.	127	259	181	4.0	6.6
	5B.	145	293	150	4.1	6.8
	6B.	146	258	168	3.9	6.5
	7B.	105	178	154	1.9	3.1
	8B.	107	98	155	1.2	2.1

In summary, glucose oxidase crystals were crosslinked with methods including single carbohydrate crosslinking, or single carbohydrate crosslinking followed by either glutaraldehyde crosslinking, tris pretreated glutaraldehyde crosslinking, a combination of lysine and glutaraldehyde crosslinking, and sequential and repeated glutaraldehyde and diaminooctane crosslinking. All of the methods produced controlled dissolution crystals with high enzymatic activity. In each case the second crosslinking reduced the activity of the crystals, but also reduced the solubility in the controlled dissolution buffer. In samples 1B through 6B, the solubility in 0.1 M NaOH was increased. In contrast, the results with sample Nos. 7A and 7B, in which the

- 61 -

second crosslinking was performed using the combination of glutaraldehyde and diaminooctane, showed reductions in the solubility of the crystals in both the controlled dissolution test and in 0.1 M NaOH.

- 5 Microscopic examination of the double crosslinked crystals revealed that all samples looked similar, with sharp shaped crystals that were yellow colored and some of them were broken. Thus, it appears the crystalline form was well preserved during the 10 crosslinking procedures.

Those of skill in the art will appreciate that variations on the methods described here can be used to produce highly insoluble carbohydrate crosslinked glycoprotein crystals. For example, 15 optimization of the pH conditions during the oxidation reaction step, manipulation of the oxidant concentration and time of reaction, and performance of the reduction reaction step will reduce the controlled dissolution properties of the crystals. Likewise, 20 alternating an optimized carbohydrate crosslinking with one or more pH optimized pretreated glutaraldehyde or other multifunctional crosslinking steps will greatly increase the insolubility, while maintaining activity.

Example 3 - Preparation of Gluconic Acid Using  
Carbohydrate Crosslinked Glucose Oxidase  
Crystals

Carbohydrate crosslinked glucose oxidase crystals, prepared as described above, are useful in the preparation of gluconic acid. Prior to use they 30 may be stored in phosphate buffer pH 6.2 containing 40% of ethanol to prevent microbial growth. The amount of enzyme in each example is measured as a volume of homogenous slurry. The slurry is first washed with

- 62 -

water on a membrane filter to remove ethanol and buffer. The carbohydrate crosslinked glucose oxidase crystals are added as washed cake from the membrane directly to a stirred batch of glucose solution to 5 start the reaction. The dry substance of the carbohydrate crosslinked glucose oxidase crystal slurry is 25 mg/ml.

The effects of aeration and stirring are studied prior to setting up the standard conditions. A 10 single batch of 1 M glucose solution is used. The batch is first aerated at a low rate (0.6 l/min) and stirred at a low speed (100 rpm). The rate of the reaction is followed until it is stable. Then, aeration and stirring are increased successively to 15 provide a variety of combinations of aeration and stirring conditions. The reaction rates at each aeration-stirring combination are recorded. The catalyzed reaction is highly limited by the availability of oxygen. Thus, the stirring and 20 aeration must be very effective. The aeration and stirring which provide the maximum reaction rate are selected for use as the standard conditions.

The gluconic acid production process is performed at different concentrations of glucose. The 25 same amount of carbohydrate crosslinked glucose oxidase crystals (500 mg dry substance) are used in each example. The process is carried out over a period of 300-600 minutes. Only the 100 g/l experiment is run to 100% conversion.

30 A glucose concentration of 100 g/l is selected as standard for comparison of the effects of the other parameters. The linear progress of the reaction in the 100 g/l experiment indicates that glucose concentration is not the limiting factor.

- 63 -

The effect of pH is studied in the range of 4 to 9. The pH of each experiment is adjusted by the set point of the titrator. No buffer is used. The reaction rate is favorable between pH 5 and 8. The  
5 rate is decreased to 50% at pH 4 and 9.

The amount of carbohydrate crosslinked glucose oxidase crystal used is varied between 125 and 1250 mg in the standard batches. As the amount of carbohydrate crosslinked glucose oxidase crystals  
10 increase from 125 to 500 mg, the reaction rate increases in direct proportion. The highest dose of carbohydrate crosslinked glucose oxidase crystals, 1250 mg, does not increase the reaction rate beyond that realized using 500 mg.

15 The effect of catalase is studied by adding either 0.0 ml, 0.1 ml or 0.3 ml to the reaction. An industrial catalase is used for comparison: Genencor CAT HP L5000, Lot No. 6094186026, date APR 11.95. Activity 5000 Baker units per ml. One unit will  
20 decompose 264 mg (7.76 millimoles) hydrogen peroxide in one hour at 25°C.

The highest dosage of catalase decomposes 194 millimoles of hydrogen peroxide per minute, which is more than 200 times the activity of glucose oxidase,  
25 0.91 mmol/min, in this experiment. However, catalase addition has no significant effect on the reaction rate. The carbohydrate crosslinked glucose oxidase crystals decompose the hydrogen peroxide which is produced in the reaction.

30 Gluconic acid production at three different temperatures (10, 25 and 30°C) are studied using standard conditions. The rate of reaction is approximately the same at 25 and 30°C, but significantly lower at 10°C. The solubility of oxygen

- 64 -

may decrease at higher temperatures and may becoming a limiting factor.

Example 4 - Preparation of Crosslinked Candida Rugosa Crystals

5       A. Crystallization of Candida Rugosa Lipase

A 5 kg aliquot of Candida rugosa lipase ("CRL") in powder form (Meito) was mixed with 5 kg celite and dissolved in 102 L distilled deionized water and the volume brought to 200 L with distilled 10 deionized water. The suspension was mixed in an Air Drive Lightning Mixer for 2 hours at room temperature and then filtered through a 0.5 micron filter to remove celite. The mixture was then ultrafiltered and concentrated to 14 L (469 g) using a 3K hollow fiber 15 filter membrane cartridge. Solid calcium acetate was added to a concentration of 5 mM Ca(CH<sub>3</sub>COO)<sub>2</sub>. The pH was adjusted to pH 5.5 with concentrated acetic acid, as necessary. A 7 liter aliquot was crystallized by either addition of 1.75 liters of 2-methyl-2,4-pentanediol ("MPD") or by addition of 3.5 liters of a 20 30% solution of PEG-8000. The resulting solution was mixed and crystallization allowed to proceed overnight at ambient temperature for about 17-20 hrs. The crystal yield was about 70%.

25       B. Carbohydrate Crosslinking of Candida Rugosa Lipase

244 mg of enzyme crystals were suspended in 2 ml of 50 mM sodium acetate, 5 mM CaCl<sub>2</sub>, and 10% polyethylene glycol at pH 5.5. The crystals were then 30 oxidized with 2-15 mg of sodium periodate (NaIO<sub>4</sub>) for 2 hours at room temperature and overnight at 0°C. Oxidation was followed by dialysis in the 50 mM sodium

- 65 -

acetate, 5 mM CaCl<sub>2</sub>, and 10% polyethylene glycol at pH 5.5 to remove periodate. The oxidized crystals were exposed to 200  $\mu$ L of an 0.8 M solution of ethylenediamine solution in 50 mM Tris, 5 mM CaCl<sub>2</sub> at 5 pH 8.0 and the reaction was allowed to proceed for 2 hours. The mixture was acidified to pH 6.0 using 1 N HCl and then 20  $\mu$ l of 0.1 M NaCNBH<sub>3</sub> was added at the end of 2 hours. The reaction was allowed to proceed for another 30 minutes. The protein was then washed 10 several times with 10 mM Tris, 10 mM CaCl<sub>2</sub> at pH 6.0 and suspended in the same buffer. The modified samples were insoluble in 10 mM Tris, 10 mM CaCl<sub>2</sub> at pH between 6 and 7, but were soluble when the pH was raised to 8.5 or higher. They retained a high level of original 15 activity towards p-nitrophenyl acetate. The activity was followed spectrophotometrically at 410 nm for the release of p-nitrophenol. The assay solution had 1 mM of p-nitrophenylacetate in 50 mM sodium acetate buffer at pH 6.5. The extinction coefficient of p-nitrophenol 20 at this pH is 2,500 M<sup>-1</sup>cm<sup>-1</sup>.

It should be understood that the solubility of the protein may be efficiently controlled at each stage of the above-described process by varying the reaction conditions to achieve a different extent or 25 degree of oxidation and by changing the stoichiometry and hydrophobicity of the diamine component (e.g., changing from ethylene diamine to hexane diamine).

In one example, 875 mg of Candida rugosa lipase in 35 ml of 0.05 M acetate, 10% PEG 8000 pH 5.5 30 was oxidized with 200 mg of sodium periodate for two hours at room temperature or overnight at 0 °C. After oxidation, the crystals were dialyzed in 10 mM Tris, 10% PEG 8000 to remove the periodate. The oxidized crystals were then incubated with 14 mls of a 1 M

- 66 -

solution hexanediamine hydrochloride at pH 8 and 315 µl of 1 N NaOH was added to the dialyzed enzyme and Schiff-base formation was allowed to proceed for 6 hrs at room temperature. The reaction mix was acidified to 5 pH 6 with 1 N HCl, requiring approximately 400 µl of 1 N HCl and 400 mg of solid NaCNBH<sub>3</sub>, was added. The reduction reaction was allowed to proceed for 90 minutes at room temperature. Next the mixture was centrifuged and washed 4 times with 10 mM CaCl<sub>2</sub>. The 10 crosslinked enzyme was stored in 10 mM Tris, 10 mM CaCl<sub>2</sub> at pH 7.0.

This crosslinked sample can be further modified with glutaraldehyde and/or other reagents as shown below. Further modifications by glutaraldehyde 15 and other reagents may occur either at the amino groups introduced at the surface or possibly at amino groups within amino acid side chains.

In another example, 75 mg of the crosslinked sample from above was suspended in 0.5 ml of 10 mM 20 CaCl<sub>2</sub> at pH 7.0. Next 50 µl of a 25 % pretreated glutaraldehyde solution was added and the reaction was allowed to proceed for 18 hours at room temperature. Pretreated glutaraldehyde was prepared by mixing 1:1 of 25 50% glutaraldehyde with 0.3 M Borate buffer at pH 8.5 and heating at 50 °C for 1hour. The reaction was terminated by centrifugation of the slurry, followed by decanting of the supernatant, and washing of the glycoprotein crystals four times with 10 mM Tris, 10 mM CaCl<sub>2</sub> at pH 7.0.

30 Two different simultaneous surface modifications can be introduced onto the oxidized glycoprotein crystals using two different crosslinking reagents. This entails simultaneous addition of pretreated glutaraldehyde (12 µl of a 25 % solution)

- 67 -

and 2 % (w/v) of a second reagent which was either succinic anhydride, phenylglutaric anhydride, salicylaldehyde, acetimidate, formalin, acrolein, succinic semialdehyde, butyraldehyde, dodecylaldehyde, 5 glyceraldehyde or trans-oct-2-enal. to 75 mg of crystals in 0.5 ml of 10 mM CaCl<sub>2</sub> at pH 7.0. All reactions were carried out at pH 7.5-8.0 for twenty four hours at room temperature with shaking. The reaction was terminated by centrifugation of the 10 crystals, decanting of the supernatant and subsequently followed by five washes with 10 mM CaCl<sub>2</sub> at pH 7.0.

All of the samples of *Candida rugosa* lipase crosslinked via the carbohydrate method were insoluble and did not leach protein after a 24 hr incubation 15 period at 40 °C in 10 mM Tris, 10 mM CaCl<sub>2</sub> at pH 7.0.

Example 5

Retrovirus Vaccine Using Carbohydrate Crosslinked Crystals of Viral Proteins

A vaccine comprising an antigen component and 20 an adjuvant component for immunopotentiation is prepared as follows. A retroviral vaccine antigen is constructed using carbohydrate crosslinked glycoprotein crystals of HIV gp160, gp120, gp41, or co-crystals of fragments of the protein complexed with CD4 or 25 antibodies, prepared according to the above-described methods. Co-crystallization of gp120 and an antibody to the CD4 binding region may be necessary to expose the chemokine receptor binding sites (See R. Wyatt and J. Sodroski, "The HIV-1 envelope glycoproteins: 30 Fusogens, Antigens, and Immunogens," *Science* 280:1884-1888 (1998)). Many genetic clones of both primary and laboratory isolates of HIV are available from the Aids

- 68 -

Research and Reagent Program, National Institutes of Allergy and Infectious Diseases, Rockville MD 20852, for designing vaccines which induce broad neutralizing immunity. Alternatively, protein from an attenuated 5 HIV-1 or HIV-2 strain of virus may be chosen to construct a vaccine. In an alternate embodiment, carbohydrate crosslinked glycoprotein crystals from various isolates or strains of virus may be combined to broaden the scope of immunity. In addition, specific 10 synthetic or recombinant fragments of virus protein may also be used as the antigen component of the vaccine.

One advantage of the present invention is that antigen presentation of the protein epitopes can be optimized by using only carbohydrate crosslinked 15 glycoprotein crystals. In addition, the antigen component of the vaccine can be a mixture of crystals, in which a subset of crystals are crosslinked primarily through the amino acid side chains with little or no carbohydrate crosslinking. In this way, antigen 20 presentation of native or unmodified carbohydrate antigens may help to optimize the immune response. Various combinations of crystals derived from proteins and glycoproteins including gp160, gp120, gp41, p26, or co-crystals of fragments of the glycoprotein complexed 25 with CD4 or antibodies, viral structural proteins, viral enzymes, and viral regulatory/accessory proteins, are prepared using different specific crosslinking treatments (protein and carbohydrate specific) and mixed in the vaccine to induce the broadest possible 30 neutralizing antibody and T-cell response. See M. Emerman and M.H. Malin "HIV-1 Regulatory/Accessory genes: Keys to Unraveling Viral and Host Cell Biology," Science 280: 1880-1884 (1998); N.L. Letvin, "Progress

- 69 -

in the Development of an HIV-1 Vaccine," *Science* 280: 1875-1880 (1998); and R. Wyatt and J. Sodroski, "The HIV-1 envelope glycoproteins: Fusogens, Antigens, and Immunogens," *Science* 280: 1884-1888 (1998).

5 An advantage of the carbohydrate crosslinked glycoprotein crystals of this invention is that they can be maintained in buffer, formulated directly with adjuvants, or dried for storage with subsequent reformulation and administration at the site of  
10 immunization.

Suitable adjuvants or immunopotentiators useful in preparing vaccines according to this invention include (1) aluminum salts, such as alum; (2) surface active agents, such as Saponins and Quil A; (3) 15 polyanions; (4) bacterial products and derivatives thereof, such as Freund's complete and Freund's incomplete adjuvant, N-acetylmuramyl-L-alanyl-D-isoglutamine (muramyl dipeptides MDP), N-acetylmuramyl-L-threonyl-D-isoglutamine (threonyl MDP), Lipid A, 20 bacterial lipopolysaccharides; (5) vehicles and slow release materials such as liposomes; and (6) cytokines such as IL-2 and IFN- $\gamma$ .

The dosage amount for such vaccines and frequency of boosting will depend on the age and weight 25 of the person or animal being immunized and the immunogenicity of the vaccine preparation. Methods for formulating, administering, conducting clinical trials for safety and efficacy of vaccine candidates are known to those of skill in the art (See *New Generation Vaccines*, Edited by G.C. Woodrow and M.M. Levine, 30 (1990) Marcel Decker, Inc. New York, NY). The immune response and vaccine is assessed by the level and specificity of the antibody and T-cell responses to

- 70 -

viral proteins and peptides. Techniques for measuring antibody and T-cell responses to HIV proteins and glycoproteins are well known in the art (See N.L. Letvin, "Progress in the Development of an HIV-1 5 Vaccine," *Science* 280: 1875-1880 (1998), and references cited therein).

With the HIV-1/AIDS vaccine as one example, those of skill in the art will appreciate that the present invention is usefull to provide vaccines for 10 other diseases or in circumstances in which improvement of an existing vaccine is desired. For example, vaccine applications for the present invention include the following diseases: (1) viral diseases such as cytomegalovirus, herpes simplex, genital herpes, 15 Hepatitis A, Non A Non B hepatitis, parainfluenza, respiratory syncytial virus, rotavirus, and lassa fever; (2) bacterial diseases such as leprosy, gonorrhea, enterotoxigenic *E. coli*, *streptococcus pyogenes*, *streptococcus mutans*, *klebsiella*, and 20 *pseudomonas*; and (3) Parasitic diseases such as *Ascaris*, Malaria, Amebae, Schistosomes, Arypanosomes, Hookworm, Trichuris, Filaris, Giardia, and Leishmania.

While we have hereinbefore described a number of embodiments of this invention, it is apparent that 25 our basic constructions can be altered to provide other embodiments which utilize the processes and compositions of this invention. Therefore, it will be appreciated that the scope of this invention is to be defined by the claims appended hereto rather than by 30 the specific embodiments which have been presented hereinbefore by way of example.

- 71 -

CLAIMS

We Claim:

1. A crosslinked glycoprotein crystal, said crystal being crosslinked through one or more carbohydrate moieties in said glycoprotein.

2. The crosslinked glycoprotein crystal according to claim 1, wherein said crystal is crosslinked with a multifunctional crosslinking agent.

3. The crosslinked glycoprotein crystal according to claim 1, wherein said crystal is crosslinked with a diamine crosslinking agent.

4. The crosslinked glycoprotein crystal according to claim 3, wherein said diamine crosslinking agent is selected from the group consisting of hexamethylenediamine, diaminooctane, and ethylenediamine.

5. A crosslinked glycoprotein crystal, said crystal being crosslinked through one or more carbohydrate moieties in said glycoprotein and through one or more amino acid side chain functional groups in said glycoprotein.

6. The crosslinked glycoprotein crystal according to claim 5, wherein said crystal is crosslinked through one or more carbohydrate moieties in said glycoprotein with a diamine crosslinking agent and said crystal is crosslinked through one or more

- 72 -

amino acid side chain functional groups in said glycoprotein with a multifunctional crosslinking agent.

7. The crosslinked glycoprotein crystal according to claim 6, wherein said diamine crosslinking agent is selected from the group consisting of hexamethylenediamine and diaminooctane and wherein said multifunctional crosslinking agent is selected from the group consisting of glutaraldehyde, succinic anhydride, phenylglutaric anhydride, salicylaldehyde, acetimidate, formalin, acrolein, succinic semialdehyde, butyraldehyde, dodecylaldehyde, glyceraldehyde or trans-oct-2-enal.

8. The crosslinked glycoprotein crystal according to claim 1 or 5, said glycoprotein crystal having at least about 2 fold greater stability as measured by pronase digestion, thermal stability, or activity in 50% ethanol than the soluble form of the glycoprotein that is crystallized to form said glycoprotein crystal that is crosslinked.

9. The crosslinked glycoprotein crystal according to claim 1 or 5, said glycoprotein crystal having greater stability at an alkaline pH or an acidic pH than the soluble form of the glycoprotein that is crystallized to form said glycoprotein crystal that is crosslinked.

10. The crosslinked glycoprotein crystal according to claim 1 or 5, said glycoprotein crystal having greater stability at high temperature than the soluble form of the glycoprotein that is crystallized to form said glycoprotein crystal that is crosslinked.

- 73 -

11. The crosslinked glycoprotein crystal according to claim 1 or 5, said glycoprotein crystal having activity in an organic solvent or aqueous/organic solvent mixture that is at least about 2 times greater than the activity of an equivalent amount of the soluble form of the glycoprotein that is crystallized to form said glycoprotein crystal that is crosslinked.

12. The crosslinked glycoprotein crystal according to claim 8, wherein the stability of said glycoprotein crystal is at least about 5 to about 10 times that of the soluble form of the glycoprotein that is crystallized to form said glycoprotein crystal that is crosslinked.

13. The crosslinked glycoprotein crystal according to claim 1, said glycoprotein crystal being substantially insoluble and stable in a composition under storage conditions and substantially soluble and active under conditions of use of said composition.

14. The crosslinked glycoprotein crystal according to claim 13, wherein said composition is selected from the group consisting of pharmaceuticals, vaccines, immunotherapeutics, immune modifiers, adjuvants, veterinary pharmaceuticals and vaccines, agricultural compounds, foods, feeds, cosmetics, detergents, and decontamination agents.

15. The crosslinked glycoprotein crystal according to claim 1, said glycoprotein crystal being capable of releasing its protein activity at a controlled rate upon exposure to a change in the

- 74 -

environment surrounding said crystal, said change being selected from the group consisting of change in pH, change in solute concentration, change in temperature, change in chemical composition, change in shear force acting upon the crystals and combinations thereof.

16. The crosslinked glycoprotein crystal according to claim 1 or 5, wherein said glycoprotein is selected from the group consisting of hormones, enzymes, antibodies, viral receptors, viral surface glycoproteins, parasite glycoproteins, parasite receptors, T-cell receptors, MHC molecules, immune modifiers, tumor antigens, mucins, inhibitors, growth factors, trophic factors, cytokines, lymphokines, cytokines, toxoids, nerve growth hormones, blood clotting factors, adhesion molecules, multidrug resistance proteins, therapeutic proteins, adenylylate cyclases, bone morphogenic proteins and lectins.

17. The crosslinked glycoprotein crystal according to claim 16, wherein said enzyme is selected from the group consisting of hydrolases, isomerases, lyases, ligases, transferases and oxidoreductases.

18. The crosslinked glycoprotein crystal according to claim 17, wherein said enzyme is selected from the group consisting of lipases and oxidases.

19. The crosslinked glycoprotein crystal according to claim 17, wherein said enzyme is glucose oxidase.

20. The crosslinked glycoprotein crystal according to claim 17, wherein said enzyme is Candida rugosa lipase.

- 75 -

21. The crosslinked glycoprotein crystal according to claim 1 or 5, said crystal having a longest dimension of between about 0.01  $\mu\text{m}$  and about 500  $\mu\text{m}$ .

22. The crosslinked glycoprotein crystal according to claim 21, said crystal having a longest dimension of between about 0.1  $\mu\text{m}$  and about 50  $\mu\text{m}$ .

23. The crosslinked glycoprotein crystal according to claim 1 or 5, said crystal having a shape selected from the group consisting of spheres, needles, rods, plates, rhomboids, cubes, squares, bipryamids and prisms.

24. A method for producing a crosslinked glycoprotein crystal comprising the steps of:

(a) reacting a glycoprotein crystal with an oxidizing agent to oxidize the carbohydrate moieties of said glycoprotein; and

(b) reacting said glycoprotein crystal with a crosslinking agent, under conditions sufficient to induce crosslinking of said crystal through the carbohydrate moieties of said glycoprotein.

25. The method for producing a crosslinked glycoprotein crystal according to claim 24, wherein said crosslinking agent is a multifunctional crosslinking agent.

26. The method for producing a crosslinked glycoprotein crystal according to claim 24, wherein said crosslinking agent is a diamine crosslinking agent.

- 76 -

27. The method for producing a crosslinked glycoprotein crystal according to claim 26, wherein said diamine crosslinking agent is selected from the group consisting of hexamethylenediamine, diaminoctane, and ethylenediamine.

28. The method for producing a crosslinked glycoprotein crystal according to claim 24, wherein said oxidizing agent is sodium periodate ( $\text{NaIO}_4$ ).

29. A method for producing a crosslinked glycoprotein crystal comprising the step of reacting a glycoprotein crystal with a first crosslinking agent, said first crosslinking agent being capable of crosslinking said crystal through the amino acid side chain functional groups of said glycoprotein and at least a second crosslinking agent, said second crosslinking agent being capable of crosslinking said glycoprotein through carbohydrate moieties of said glycoprotein.

30. The method according to claim 29, wherein said glycoprotein crystal is crosslinked with said first and said second crosslinking agent at the same time or in sequence.

31. A method for producing a crosslinked glycoprotein crystal comprising the steps of:

(a) reacting a glycoprotein crystal with a first crosslinking agent, under conditions sufficient to induce crosslinking through one or more carbohydrate moieties of said glycoprotein; and

- 77 -

(b) reacting said crosslinked glycoprotein crystal with at least a second crosslinking agent, under conditions sufficient to induce crosslinking of said crystal through one or more carbohydrate moieties of said glycoprotein, said second crosslinking agent being different from said first crosslinking agent.

32. A method for producing a crosslinked glycoprotein crystal comprising the steps of:

(a) reacting a glycoprotein crystal with a first crosslinking agent, under conditions sufficient to induce crosslinking through one or more amino acid side chain functional groups of said glycoprotein; and

(b) reacting said crosslinked glycoprotein crystal with least a second crosslinking agent, under conditions sufficient to induce crosslinking of said crystal through one or more carbohydrate moieties of said glycoprotein.

33. A method for producing a crosslinked glycoprotein crystal comprising the steps of:

(a) reacting a glycoprotein crystal with a first crosslinking agent, under conditions sufficient to induce crosslinking through one or more amino acid side chain functional groups of said glycoprotein; and

(b) reacting said crosslinked glycoprotein crystal with a second crosslinking agent and at least a third crosslinking agent, said second crosslinking agent being capable of crosslinking said glycoprotein through one or more amino acid side chain functional groups of said glycoprotein and said third crosslinking agent being capable of crosslinking said glycoprotein through one or more carbohydrate moieties of said glycoprotein.

- 78 -

34. A method for producing a crosslinked glycoprotein crystal comprising the steps of:

(a) reacting a glycoprotein crystal with a first crosslinking agent, under conditions sufficient to induce crosslinking through one or more carbohydrate moieties of said glycoprotein; and

(b) reacting said crosslinked glycoprotein crystal with a second crosslinking agent and at least a third crosslinking agent, said second crosslinking agent being capable of crosslinking said glycoprotein through one or more amino acid side chain functional groups of said glycoprotein and said third crosslinking agent being capable of crosslinking said glycoprotein through one or more carbohydrate moieties of said glycoprotein.

35. The method for producing a crosslinked glycoprotein crystal according to any one of claims 29, 31, 32, 33, or 34, wherein said crosslinking agent capable of crosslinking said glycoprotein through the carbohydrate moieties of said glycoprotein is a multifunctional crosslinking agent.

36. The method for producing a crosslinked glycoprotein crystal according to any one of claims 29, 31, 32, 33, or 34 wherein said crosslinking agent capable of crosslinking said glycoprotein through the carbohydrate moieties of said glycoprotein is a diamine crosslinking agent.

37. The method for producing a crosslinked glycoprotein crystal according to claim 36, wherein said diamine crosslinking agent is selected from the group consisting of hexamethylenediamine and diaminooctane, and ethylenediamine.

- 79 -

38. The method for producing a crosslinked glycoprotein crystal according to any one of claims 29, 31, 32, 33, or 34, wherein said crosslinking agent capable of crosslinking said glycoprotein through the amino acid side chains of said glycoprotein is a multifunctional crosslinking agent.

39. The method for producing a crosslinked glycoprotein crystal according to claim 38, wherein said multifunctional crosslinking agent is selected from the group consisting of glutaraldehyde, succinic anhydride, phenylglutaric anhydride, salicylaldehyde, acetimidate, formalin, acrolein, succinic semialdehyde, butyraldehyde, dodecylaldehyde, glyceraldehyde or trans-oct-2-enal.

40. The method for producing a crosslinked glycoprotein crystal according to any one of claims 29, 31, 32, 33, or 34, wherein said glycoprotein is selected from the group consisting of hormones, enzymes, antibodies, viral receptors, viral surface glycoproteins, parasite glycoproteins, parasite receptors, T-cell receptors, MHC molecules, immune modifiers, tumor antigens, mucins, inhibitors, growth factors, trophic factors, cytokines, lymphokines, cytokines, toxoids, nerve growth hormones, blood clotting factors, adhesion molecules, multidrug resistance proteins, therapeutic glycoproteins, adenylate cyclases, bone morphogenic proteins and lectins.

41. The method according to claim 40, wherein said enzyme is selected from the group consisting of hydrolases, isomerases, lyases, ligases, transferases and oxidoreductases.

- 80 -

42. The method according to claim 41,  
wherein said enzyme is selected from the group  
consisting of lipases and oxidases.

43. The method according to claim 41,  
wherein said enzyme is glucose oxidase.

44. The method according to claim 41,  
wherein said enzyme is Candida rugosa lipase.

45. The method for producing a crosslinked  
glycoprotein crystal according to any one of claims 29,  
31, 32, 33, or 34, wherein, prior to reacting said  
glycoprotein crystal with said crosslinking agent, said  
method further comprises the step of crystallizing said  
glycoprotein.

46. The method according to claim 45,  
wherein the step of crystallizing said glycoprotein is  
carried out without deglycosylating said glycoprotein.

47. The method according to claim 46,  
wherein the step of crystallizing said glycoprotein is  
carried out in the presence of a water miscible solvent  
and polyethylene glycol.

48. The method for producing a crosslinked  
glycoprotein crystal according to any one of claims 29,  
31, 32, 33, or 34, wherein, prior to reacting said  
glycoprotein crystal with said crosslinking agent, said  
method further comprises the step of oxidizing the  
carbohydrate moieties of said glycoprotein.

49. A protein delivery system, said system  
comprising crosslinked glycoprotein crystals according  
to claim 15.

- 81 -

50. A pharmaceutical controlled release formulation comprising crosslinked glycoprotein crystals according to claim 15.

51. A vaccine comprising crosslinked glycoprotein crystals according to claim 1 or 5.

52. A biosensor for detecting an analyte of interest in a fluid, comprising:

(a) a crosslinked glycoprotein crystal according to claim 1 or 5, wherein said glycoprotein has the activity of acting on the analyte of interest or on a reactant in a reaction in which the analyte of interest participates;

(b) a retaining means for said crosslinked glycoprotein crystal, said retaining means consisting of a material which allows contact between said crosslinked glycoprotein crystal and a fluid, said fluid containing either (1) the analyte upon which the glycoprotein acts or (2) a reactant in a reaction in which the analyte participates; and

(c) a signal transducer which produces a signal in the presence of the analyte.

53. A method for detecting the presence of a substance in a sample comprising the steps of:

(a) contacting the sample with a crosslinked glycoprotein crystal according to claim 1 or 5, wherein said glycoprotein has the activity of acting on the substance or on a reactant in a reaction in which the substance participates to cause a detectable change, under conditions and for a time sufficient to permit said activity; and

(b) using a detecting means to detect the detectable change.

- 82 -

54. A method for producing a selected product using a glycoprotein comprising the steps of:

- (a) combining at least one substrate with at least one glycoprotein which acts upon the substrate, said glycoprotein being a crosslinked glycoprotein crystal according to claim 1 or 5; and
- (b) maintaining the combination produced in step (a) under conditions which permit said glycoprotein to act upon the substrate, thereby producing the selected product.

55. A method for crystallizing a glycoprotein without removing the carbohydrate moieties of said glycoprotein comprising the steps of:

- (a) concentrating pure glycoprotein to produce glycoprotein concentrate;
- (b) contacting said glycoprotein concentrate with polyethylene glycol to produce glycoprotein polyethylene glycol concentrate;
- (c) contacting said glycoprotein polyethylene glycol concentrate with a miscible organic solvent to produce a glycoprotein polyethylene glycol organic solvent concentrate;
- (d) contacting said glycoprotein polyethylene glycol organic solvent concentrate with an acid to adjust the pH to between 4.0 and 5.0; and
- (e) cooling the pH adjusted concentrate to a temperature between 4 and 6 °C for a period of time sufficient to induce crystallization.

## INTERNATIONAL SEARCH REPORT

Int. Application No

PCT/US 98/16372

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 C07K9/00 C07K14/00 C07K16/00 C07K17/00 C12N11/00  
 C12N9/96 C12N9/04 C12N9/18 A61K39/00 A61K38/00  
 A61K9/00 C1201/00 C12P1/00 C12M1/40 //C12P7/58,

According to International Patent Classification(IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 C07K C12N A61K C12Q C12P

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	A.L. MARGOLIN: "NOVEL CRYSTALLINE CATALYSTS" TRENDS IN BIOTECHNOLOGY., vol. 14, no. 7, July 1996, pages 223-230, XP004035759 CAMBRIDGE GB see the whole document	1-55
A	J.J. LALONDE ET AL.: "CROSS-LINKED CRYSTALS OF CANDIDA RUGOSA LIPASE: HIGHLY EFFICIENT CATALYSTS FOR THE RESOLUTION OF CHIRAL ESTERS." JOURNAL OF THE AMERICAN CHEMICAL SOCIETY., vol. 117, no. 26, 5 July 1995, pages 6845-6852, XP002041053 WASHINGTON, D.C., US cited in the application see the whole document ---	1-55 -/-

 Further documents are listed in the continuation of box C. Patent family members are listed in annex.

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Date of the actual completion of the international search

13 November 1998

Date of mailing of the international search report

27/11/1998

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# INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 98/16372

**A. CLASSIFICATION OF SUBJECT MATTER**

IPC 6 A61K39/21

According to International Patent Classification (IPC) or to both national classification and IPC

**B. FIELDS SEARCHED**

Minimum documentation searched (classification system followed by classification symbols)

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

**C. DOCUMENTS CONSIDERED TO BE RELEVANT**

Category <sup>a</sup>	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	WO 92 02617 A (VERTEX PHARMACEUTICALS INCORPORATED) 20 February 1992 see claims 14-24 ---	1-55
A	B. KOZULIC' ET AL.: "STUDY OF THE CARBOHYDRATE PART OF YEAST ACID PHOSPHATASE." BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 122, no. 3, 1984, pages 1083-1090, XP002084361 ORLANDO, FL US see page 1085, line 3 - line 13 see page 1089, paragraph 4 - paragraph 5 ---	1-55
A	GB 2 225 324 A (B. KOZULIC' ET AL.) 30 May 1990 see page 4, paragraph 1; claims ---	1-55

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Date of the actual completion of the international search	Date of mailing of the international search report
13 November 1998	
Name and mailing address of the ISA European Patent Office, P.B. 5818 Patentlaan 2 NL - 2280 HV Rijswijk Tel. (+31-70) 340-2040, Tx. 31 651 epo nl. Fax: (+31-70) 340-3016	Authorized officer  Ryckebosch, A

**INTERNATIONAL SEARCH REPORT**

International Application No

PCT/US 98/16372

**C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT**

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,A	WO 97 44445 A (ALTUS BIOLOGICS INC.) 27 November 1997 see page 11, line 19 - page 18, line 7; claims -----	1-55

# INTERNATIONAL SEARCH REPORT

Information on patent family members

International Application No

PCT/US 98/16372

Patent document cited in search report		Publication date		Patent family member(s)		Publication date
WO 9202617	A	20-02-1992		AU 662104 B AU 8315491 A CA 2087730 A EP 0550450 A EP 0861888 A FI 930454 A HU 65904 A HU 9500666 A LT 1077 A,B LV 10304 A,B NZ 264042 A PT 98564 A SK 5993 A US 5618710 A US 5801022 A		24-08-1995 02-03-1992 04-02-1992 14-07-1993 02-09-1998 02-02-1993 28-07-1994 28-11-1995 25-04-1995 20-10-1994 25-06-1996 30-06-1992 07-07-1993 08-04-1997 01-09-1998
GB 2225324	A	30-05-1990		US 5258501 A		02-11-1993
WO 9744445	A	27-11-1997		AU 3072897 A		09-12-1997